

Studies of pharmacokinetic and pharmacodynamic properties of isoallopregnanolone in healthy women

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Abstract

Rationale The pharmacokinetics and behavioral effects of isoallopregnanolone (3 β -hydroxy-5 α -pregnan-20-one) in women are not known.

Objectives Allopregnanolone (3 α -hydroxy-5 α -pregnan-20-one) is a well-known neurosteroid, acting via the GABA_A receptor in the human brain. The naturally occurring progesterone metabolite isoallopregnanolone is the 3 β -stereoisomer of allopregnanolone. Prior studies have concluded that isoallopregnanolone has no effect on the GABA_A receptor. However, an antagonistic effect of isoallopregnanolone to allopregnanolone on the GABA_A receptor has been shown in animal and in vitro studies. The purpose of this study was to evaluate the pharmacokinetics and behavioral effects of isoallopregnanolone in humans.

Materials and methods Six healthy women were given three increasing doses of isoallopregnanolone intravenously in the follicular phase. Repeated blood samples for analyses of isoallopregnanolone and allopregnanolone concentrations were drawn. Saccadic eye movement variables, self-rated sedation, and mood rating scales were used during the test day. A Likert scale for prospective symptoms was used to measure daily fluctuations during the ongoing menstrual cycle.

Results Exogenously administered isoallopregnanolone produced a dose-dependent increase in the serum concentration of isoallopregnanolone. In parallel, there was also a rise in the allopregnanolone concentration. There was a decrease in saccadic eye movement variables, but no effect was found on self-rated sedation or mood and no changes were seen in prospective symptoms during the menstrual cycle.

Conclusions After administration of isoallopregnanolone at a cumulative dose of 0.20 mg/kg, no adverse effects were observed. There is a metabolism of isoallopregnanolone to allopregnanolone, most likely explaining the effects on the saccadic eye movements.

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Introduction

Neuroactive steroids, especially metabolites of progesterone, are potent modulators of the gamma-aminobutyric acid A (GABA_A) receptor and exercise rapid effects on brain function (Majewska et al. 1986; Baulieu 1991). Allopregnanolone (3 α -hydroxy-5 α -pregnan-20-one) is one of the

most potent and well-known neuroactive steroids. However, its stereoisomer, isoallopregnanolone (3 β -hydroxy-5 α -pregnan-20-one), does not seem to have any effect on the GABA system by itself (Lan et al. 1991; Stromberg et al. 2006).

Allopregnanolone has agonistic effects on the GABA_A receptor complex in a way similar to other GABA agonists like benzodiazepines, barbiturates, and alcohol. Allopregnanolone potentiates the effect of GABA on the receptor by lengthening the opening time of the chloride channel and by increasing the frequency of channel openings, which are actions similar to both barbiturates and benzodiazepines (Belelli and Lambert 2005). Through this effect, allopregnanolone enhances inhibitory neurotransmission (Majewska et al. 1986), thus exerting sedative (Lancel et al. 1997; Timby et al. 2006) and antiepileptic effects (Lundgren et al. 1998).

Isoallopregnanolone is a steroid closely related to allopregnanolone. The only structural difference from its isomer allopregnanolone is the hydroxyl group in 3 β -instead of 3 α -position. Isoallopregnanolone by its own is, as far as we know today, without hormonal or GABA_A receptor effects (Lundgren et al. 2003; Stromberg et al. 2006). Instead, isoallopregnanolone has often been used as a control when testing the specificity of allopregnanolone and its GABA_A-receptor-enhancing effect to show that the sterical configuration with the 3-hydroxy group is of major importance for the GABA_A-receptor-stimulating effects of allopregnanolone (Gee et al. 1987). Furthermore, no isoallopregnanolone influence has been noted on anesthesia (Gyermek et al. 1968), anxiolysis (Bitran et al. 1991; Wieland et al. 1991; Carboni et al. 1996; Rodgers and Johnson 1998), antiepileptic effect (Kokate et al. 1994), hyperphagic effect (Chen et al. 1996), or stimulation of gastric acid secretion (Watanabe et al. 2000). Also, in *in vitro* experiments, no effect of isoallopregnanolone of its own has been detected (Peters et al. 1988; Lan et al. 1991; Lambert et al. 1995; Dayanithi and Tapia-Arancibia 1996; Le Foll et al. 1997; Calogero et al. 1998; Concas et al. 1998; Lundgren et al. 2003).

However, it has recently been shown that isoallopregnanolone can antagonize the effect of allopregnanolone on the GABA_A receptor as shown *in vitro* (Wang et al. 2002; Lundgren et al. 2003) on slices of brain tissue (Wang et al. 2000) and *in vivo* as it inhibits allopregnanolone-mediated induction of anesthesia in rats (Backstrom et al. 2005). The antagonizing effect of isoallopregnanolone on allopregnanolone seems specific as GABA, benzodiazepine, or barbiturate effects are not inhibited by isoallopregnanolone (Lundgren et al. 2003; Stromberg et al. 2006).

Isoallopregnanolone is synthesized in the mammalian ovary and the serum concentration varies over the menstrual cycle and in pregnancy in parallel to progesterone

and allopregnanolone (Coperchot et al. 1993; Parizek et al. 2005; Havlikova et al. 2006; Hill et al. 2007). Within the ovary, allopregnanolone originates predominantly from the corpus luteum (Ottander et al. 2005). In addition, isoallopregnanolone and allopregnanolone are synthesized from the adrenal cortex and a small amount of allopregnanolone and isoallopregnanolone can be synthesized *de novo* in the central nervous system from cholesterol (Baulieu and Robel 1990; Coperchot et al. 1993).

Allopregnanolone seems to be involved in several clinical conditions in both women and men. Numerous studies have reported relationships between the plasma levels of progesterone–allopregnanolone and the occurrence of epileptic seizures, mood symptoms, and sedation (Backstrom 1976; Backstrom et al. 2003a; Timby et al. 2006). Allopregnanolone presence or concentrations has been studied in women in psychiatric conditions such as premenstrual dysphoric disorder (PMDD; Rapkin et al. 1997; Bicikova et al. 1998; Monteleone et al. 2000; Girdler et al. 2001; Backstrom et al. 2003b; Nyberg et al. 2007), mood symptoms during hormone therapy in the climacteric (Andreen et al. 2006), and postpartum depression (Nappi et al. 2001; Epperson et al. 2006). Altered allopregnanolone concentrations have been found in both women and in males in psychiatric conditions such as major depression and anxiety disorders (Romeo et al. 1998; Strohle et al. 2002).

In most of the menstrual cycle linked and hormone therapy studies, allopregnanolone is related to increased anxiety and thus seems to have an anxiogenic effect as there is a close relationship between increase in allopregnanolone concentrations and the development of negative mood symptoms (Backstrom et al. 2003a; Andreen et al. 2006). The anxiogenic effect is surprising but this paradoxical effect of allopregnanolone is similar to what has been noted in certain sensitive individuals with any kind of GABA_A receptor agonist like benzodiazepines (Wenzel et al. 2002), barbiturates (Masia et al. 2000), and alcohol (Miczek et al. 1997). A paradoxical effect of allopregnanolone has also been shown in animal studies (Beauchamp et al. 2000; Miczek et al. 2003) and a relation exists between allopregnanolone levels and negative mood in postmenopausal women receiving progesterone treatment (Andreen et al. 2006). In the conditions where allopregnanolone has an anxiogenic effect, treatment with an antagonistic substance would most likely be beneficial. Isoallopregnanolone could constitute such a therapy if it acts in a similar way in humans as it does in animals.

An objective measure of sedative effect is saccadic eye movement measurements. A saccade is a rapid, jump-like movement of the eye from one fixation point to another, used by the eye to change the focus of the fovea. Maximal saccadic eye velocity (SEV) has a variation between subjects (Hommer et al. 1986; Sundstrom and Backstrom

1998) but is stable within subjects, both within a testing period and between testings (Gentles and Thomas 1971; Glue et al. 1991; Hommer et al. 1986; Roy-Byrne et al. 1990; Sundstrom and Backstrom 1998). Once a saccade has started, it is outside conscious control and not subjected to motivational influences (Gentles and Thomas 1971), and therefore SEV is considered to provide an objective and sensitive measure of sedation. SEV is reduced in a dose-dependent manner by benzodiazepines, pregnanolone (3 α -hydroxy-5 β -pregnan-20-one), alcohol, and allopregnanolone (Hommer et al. 1986; Ball et al. 1991; Sundstrom et al. 1997, 1998; Nyberg et al. 2004; Timby et al. 2006). Furthermore, benzodiazepine-, pregnanolone-, and allopregnanolone-induced increase in self-ratings of sedation, which is another pharmacological action of these compounds, is highly correlated with a reduction in SEV (Hommer et al. 1986; Ball et al. 1991; Sundstrom et al. 1998; Timby et al. 2006).

Decreased SEV and increased self-rated sedation are documented effects of allopregnanolone (Timby et al. 2006) and are suitable end point variables to study. Thus, it is reasonable to study also the effects of isoallopregnanolone on sedation and SEV. As isoallopregnanolone is given to humans for the first time and it is a metabolite of progesterone it is also of interest to investigate if any other effects appear during or after the isoallopregnanolone administration. Furthermore, the pharmacokinetic properties of isoallopregnanolone given in supraphysiological doses are not known.

The aim of the present study was to determine the safety and pharmacokinetic properties of isoallopregnanolone and to investigate whether there is an effect on saccadic eye movement variables and self-rated sedation in healthy women.

Material and methods

Subjects

Seven healthy women aged 18 to 40 years and with regular menstrual cycles were recruited through advertisement in the local newspaper and screened for inclusion in the study. The exclusion criteria were treatment with any steroid compound (including oral contraceptives and hormonal intrauterine devices) during the last 6 months prior to enrollment in the study, treatment with benzodiazepines or other psychotropic drugs during the last 3 months preceding inclusion, and treatment with any drug (including over-the-counter drugs) during the last 4 weeks before inclusion. Women planning to become pregnant were excluded. Women with night work or women who had been traveling and experienced jet lag in the week before the study day

were also excluded. Further exclusion criteria were any current or previous somatic disease, any mental disorder, including PMDD, during the last 6 months, or a history of drug abuse or alcohol use more than six glasses of wine or beer during 1 day in the last 4 weeks before the study day. The presence of psychiatric disorders was evaluated using a structured psychiatric interview, Primary Care Evaluation of Mental Disorders (Spitzer et al. 1994), which has been validated for use in primary care settings and conforms to the criteria in Diagnostic and Statistical Manual of Mental Disorders, 4th Edition, DSM-IV (American Psychiatric Association 1994). Prior to inclusion, physical and gynecological examinations were performed, as well as routine blood chemistry screens (blood cell counts and plasma glucose, liver enzyme, creatinine, sodium, and potassium concentrations). All subjects had negative pregnancy tests in urine and normal blood chemistry screens. They gave oral and written informed consent prior to inclusion in the study. The study procedures were in accordance with ethical standards for human experimentation, established by the Declaration of Helsinki of 1975, revised in 1983. The Regional Ethical Review Board, Umeå University and the Medical Products Agency of Sweden approved the study.

Medication

Experimental medications were prepared by the Umeå University Hospital Pharmacy. The isoallopregnanolone solution was formulated with purified isoallopregnanolone (3 β -hydroxy-5 α -pregnan-20-one, UC1010, Umeocrine AB, Umeå, Sweden), 8 mg isoallopregnanolone dissolved in 100-ml albumin solution (Albumin, 200 mg/ml) using an ultrasound bath. The solution was then filtered through two sterile filters before given to the subjects. The final isoallopregnanolone concentration of each batch of solution was determined using high-performance liquid chromatography (HPLC) with ultra violet absorbance (Turkmen et al. 2004). The isoallopregnanolone concentration in the final albumin solution was 0.0736 ± 0.00807 mg/ml (mean \pm SD) isoallopregnanolone ($n=6$).

Study protocol

Isoallopregnanolone was administered in the mid-late follicular phase (days 6–13 in the menstrual cycle). To avoid interference of diurnal variations, all study patients were tested during the same time of the day. No subject was allowed to consume alcohol 24 h prior to testing. Caffeine and tobacco use was restricted throughout the study day.

In the morning of the study day, an intravenous cannula was inserted in each forearm, one for injections of isoallopregnanolone and the other for blood sampling for the analyses of isoallopregnanolone and allopregnanolone

in serum. Baseline levels of allopregnanolone and isoallopregnanolone were measured before the first injection. Three intravenous injections of isoallopregnanolone were given at 30-min intervals, using doses of 0.04, 0.06, and 0.10 mg/kg, thus giving a cumulative dose of 0.20 mg/kg. Each injection was given with an infusion rate of 2 mg/min. Blood samples for measuring the serum concentrations of isoallopregnanolone and allopregnanolone were drawn at 5, 13, 18, 35, 43, 48, 65, 73, 88, 95, 105, 115, 150, 330, 600, and 780 min and 23 and 31 h after the first isoallopregnanolone injection. SEV and self-rated sedation were measured 12 times during the experiment until 330 min after the first injection. Anxiety scale measurements were performed at baseline and continuously throughout the experiment as well as the next day. Vital functions were checked and adverse events were reported throughout the experiment until the next day.

Saccadic eye movements

Saccadic eye velocity was measured using electrooculography (EOG) with the CSGAAS5 system, fully documented elsewhere (Marshall et al. 1985; Marshall and Richens 1989). The technique has been used in our laboratory for several years (Sundstrom et al. 1997, 1998; Wihlback et al. 2001). The basic concept is to measure the difference in electric potential between the cornea and the retina. The test is performed in a quiet semilit room with the patient sitting in a comfortable chair. A pillow to support the patient's head prevented head movements. Electroencephalogram cup electrodes (Synetics AB, Stockholm, Sweden) with a small amount of electrode gel (Elefix, Nihon Kohden Europe, Rosbach, Germany) are used. After the skin had been exfoliated with Skinpure cream (Nihon Kohden), the electrodes are placed 1 cm lateral of the outer canthus of both eyes, with one common electrode in the center of the forehead. Electrode impedance are measured and confirmed to be less than 5 k Ω . The subject is instructed to watch an array of light-emitting diodes (LEDs) placed at eye level, 67 cm from the glabella. The target for the eye movements is an illuminated LED. The subject is asked to look at the illuminated LED and to move her eyes to the next target (the next illuminated LED) as that LED is turned off and the next one in the array is lit. Subjects are instructed not to anticipate targets.

The target movements take place at 1.5-s intervals. A fixed sequence of 4 \times 24 targets producing target steps of 10°, 20°, 30°, and 40° is displayed with a brief rest in between. The first four of these 24 target steps of each session are not included in the subsequent analyses, in order to allow the subject to adjust to the procedure. The EOG is DC-amplified and low-pass-filtered (-3 dB at 50 Hz) before being digitized to 12-bit resolution at a sampling

frequency of 250 Hz. A personal computer controls the target movements and digitalizes the waveform using an analog-digital converter. The 80 individual EOGs, resulting from the 4 \times 20 target steps, are stored and analyzed off-line according to the method of Marshall and Richens (1989). First, the digitalized data from each target displacement are processed to locate saccades. To avoid preemptive saccades and blinking artifacts, only saccades initiated 50 to 400 ms after target movements are included. Also, to be considered a saccade, the recorded eye movement has to display a velocity of more than 100°/s. In addition, each saccade is analyzed to determine the size of the saccade in degrees, the peak saccadic velocity, and latency from target movement to onset of saccade. Saccade accuracy is determined by comparing the actual eye position at the end of the saccade with the attempted target. SEV is further processed by plotting a velocity-saccade size curve, known as the main sequence (Baloh et al. 1975). The relationship between saccade size and peak velocity is important since it remains constant even when voluntary control of saccades is attempted. The main sequence is fitted by a quadratic equation to the peak velocity data using the calculated saccade angle as the independent variable. Carrying out the fitting procedure twice and weighing the second fit with the inverse of the square of the residuals from the first fit minimized the influence of outliers in the data. The values of peak velocity for 10°, 20°, 30°, and 40° saccades are calculated by interpolation. Saccades with amplitudes of 30° are chosen for further analyses as SEV reaches a maximum at approximately 30–35° of angular movement (Baloh et al. 1975). SEV is evaluated as an objective way to estimate sedation with a small intraindividual variation (Hommer et al. 1986). The computer program that is generating and calculating the saccades is developed by Cardiff Clinical Trials Ltd. and is well documented (Richens et al. 1993).

Visual analog ratings

A visual analog scale (VAS) was used for the test person to rate sedation and subjective feelings of alcohol-like intoxication (McCormack et al. 1988). The scale measured from 0 to a 10 cm where 0 equaled complete absence of sleepiness-feelings of intoxication and 10 represented falling asleep-feelings of heavy intoxication. Subjective ratings of sedation and intoxication were made at baseline, after each of the saccadic eye measurements, and at 600 and 780 min.

Mood rating scales

At baseline, three scales were used to measure the subject's tendency to develop anxiety and the actual anxiety level

from different aspects. The Swedish version of Spielberger's State-Trait Anxiety Inventory (STAI, state subscale), Anxiety Sensitivity Inventory (ASI), and Panic Symptom Scale (PSS) were used. The difference in anxiety level during the test was measured continuously with the State Anxiety and Discomfort Scale (SADS). STAI and PSS were also used after the trial to register anxiety symptoms or relief of anxiety during the first 24 h after the test.

STAI comprises 20 items tapping anxiety proneness and anxiety in relation to the experimental situation with a total range of scores from 20 to 80 (Spielberger 1983).

The ASI is a questionnaire comprising 16 items that express concerns about the possible aversive consequences of anxiety symptoms (Reiss et al. 1986). The Swedish version of ASI has satisfactory psychometric properties (Hellstadius 2000). Respondents indicate their degree of endorsement on five-point scales that range from 0 (very little) to 4 (very much), and total scores range from 0 to 64. The ASI manual reports a mean of 19.01 (SD=9.11) for nonpatients (Peterson and Reiss 1992). Mean score for panic disorder patients was reported to be 36.2 (Taylor et al. 1992).

PSS is a DSM-IV-derived panic symptom scale (Bradwejn et al. 1991), where subjects retrospectively rate the maximum intensity of panic symptoms on 18 items (0 = not present, 1 = mild, 2 = moderate, 3 = severe, 4 = extremely severe). The sum of intensity ratings was measured by summarizing each item score on the PSS (range 0–72).

SADS is a global measure of subjective discomfort. This scale was earlier evaluated in pharmacological tests on humans, where quick changes in anxiety levels could be detected (Radu et al. 2002). The SADS is a simple Likert self-rating scale, ranging from 0 (no discomfort) to 5 (worst imaginable discomfort). Intermediate levels were 1 (slight discomfort), 2 (moderate discomfort), 3 (severe discomfort), and 4 (very severe discomfort). The SADS is a global measure of the three aspects of anxiety, namely effect, somatic symptoms, and cognitions (Aluoja et al. 1997). SADS was measured at bedside repeatedly during the test day at 19 times.

Likert scale for prospective symptom ratings

The subjects performed daily mood ratings each evening from the first day of menstruation in the actual menstrual cycle until the next menstruation started. This scale was used to measure day-to-day fluctuations in mood and has been used in our laboratory in several prior studies. The scale comprises menstrual bleeding and 15 physical and mood symptoms relevant for the diagnosis PMDD according to DSM-IV (American Psychiatric Association 1994; Sundstrom et al. 1999a).

Assays of isoallopregnanolone and allopregnanolone

Serum concentrations of isoallopregnanolone and allopregnanolone were measured in duplicates by radioimmunoassay (RIA) after preassay diethyl ether extraction and separation with HPLC. Plasma (0.4 ml) was pipetted into a cylindrical flat-bottom glass vial of 20-ml volume, and water (0.5 ml) and diethyl ether (3.0 ml) were added. The samples were then allowed to stand on an orbital shaker for 10 min. Following the liquid-liquid extraction, the vials were transferred into an ethanol-dry ice bath. The water phase became frozen, and the ether phase was decanted and evaporated under a stream of nitrogen gas. To determine the analytical recovery of the technique, known amounts of isoallopregnanolone (750 pg/ml) were added to two extraction vials and the solvent was evaporated to dryness under a stream of nitrogen before addition of plasma. Prior to the HPLC analysis, the samples were resolved in 1,000 μ l of a 50:50 ethanol and water mixture (V/V). Recovery in the allopregnanolone assay was made by adding a known amount of radioactive allopregnanolone before the assay and the recovered amount was measured after the workup procedure.

High-Performance Liquid Chromatography

The chromatographic system consisted of a Waters 1515 Isocratic HPLC-Pump (Waters AB, Sweden), which delivered a mobile phase containing methanol and water, 60:40, V/V) at a flow rate of 1.0 ml/min. A Waters 717 plus Autosampler (Waters AB, Sollentuna, Sweden) was used for injection of standard solution and plasma samples (200 μ l) into a Symmetry C18 3.5 μ m 75 \times 4.6 mm i.d. separation column (Waters AB). Detection of retention times of standards and cross-reacting steroids was at 206 nm using a Waters 2487 Dual λ Absorbance Detector (Waters AB). The detector output was recorded on a PC-based Waters Breeze Chromatography Software (version 3.20). In the preparative HPLC, 1.5-ml fractions were symmetrically collected around the peak retention time for isoallopregnanolone or allopregnanolone; retention was found from injection of a standard sample. A Waters Fraction Collector II was used for collection of samples, for further analysis with radioimmunoassay. No cross-reacting steroids had retention times close to the collected fractions. Retention time for isoallopregnanolone (5 α -pregnan-3 β -ol-20-one) is 18.9 min and for allopregnanolone (5 α -pregnan-3 α -ol-20-one) 24.2 min. Retention times for cross-reacting steroids are: 5 α -pregnan-20 β -ol-3-one 26.7 min, 5 β -pregnan-3 α -ol-20-one 22.5 min, 5 β -pregnan-3,20-dione 19.4 min, 4-pregnen-3 α -ol-20-one 18.5 min, 5 β -pregnan-3 β -ol-20-one 17.9 min, 5 α -pregnan-3,20-dione 17.0 min, 5-pregnen-3 β -ol-20-one (pregnenolone) 14.7 min, 5 α -pregnan-3 α ,21-diol-20-one 10.1 min, preg-4-

ene-3,20-dione (progesterone) 9.4 min and 4-pregnen-20 α -ol-3-one 9.2 min. The eluted fraction for isoallopregnanolone and allopregnanolone was dried down in a Speed Vac spd 2010 system (Thermo Savant, Holbrook, USA).

Radioimmunoassay of isoallopregnanolone

For the isoallopregnanolone (3 β -hydroxy-5 α -pregnan-20-one) analysis, an antibody for pregnenolone (pregnenolone-3-monoheemisuccinate-HSA; ICN Pharmaceuticals, Inc. Orangeburg, NY, USA) was used as it also binds isoallopregnanolone. Cross-reactivity with isoallopregnanolone was 26.6%, allopregnanolone 13%, 5 α -pregnan-3,20-dione 7%, 5 β -pregnane-3 β -ol-20-one <1%, 5 β -pregnan-3 α -ol-20-one <1%, and 5 α -pregnan-3 α ,20 α -diol <1%. Radioactive steroid tracer used was [7-³H(N)]-pregnenolone purchased from NEN Life Science Products, Boston, MA, USA. The lowest detectable amount of isoallopregnanolone (two standard deviations above the blank mean) was 0.11 pmol or 35.8 pg. The recovery of the extraction including the HPLC procedure was 95% for isoallopregnanolone. Each isoallopregnanolone sample was measured in quadruples and all samples from each subject were run in the same assay. The standard curve was established by preparing duplicate tubes containing seven concentrations of unlabeled isoallopregnanolone to give a range from 0 to 2.5 pmol (0–800 pg) isoallopregnanolone (data not shown). The antiserum was used in a dilution of 1/320; the antibody solutions was prepared with [7-³H(N)]-pregnenolone 3×10^6 cpm per 32-ml solution containing 65-mM boric acid (Merck, Darmstadt, Germany) buffer, pH=8.0, bovine serum albumin 100 mg/ml (Sigma, St Louis, MO, USA), human gamma globulin solution 20 mg/ml (Octapharma, Stockholm, Sweden), and antibody in milliliter ratio of the different solutions: 30:1:1:0.010. The solution was allowed to equilibrate overnight at 8°C. Antibody solution (200 μ l) was added to all standard and sample tubes, and the mixture was allowed to stand overnight at 8°C. After the addition of 200 μ l saturated ammonium sulfate, each tube was again mixed and centrifuged at 20,000 RPM for 20 min. Thereafter, the supernatant was aliquoted into a counting vial and diluted with 3.0-ml Optiphase scintillation medium (Perkin Elmer, Shelton, USA). The samples were counted in a RackBeta (Wallac, Turku, Finland) scintillation counter. The sensitivity of the assays was 0.08 pmol (25 pg), with an intra-assay coefficient of variation of 8.5% and an interassay coefficient of variation of 10.7%.

Radioimmunoassay of allopregnanolone

The samples were analyzed using a polyclonal rabbit antiserum (a kind gift from RH Purdy, The Scripps

Research Institute La Jolla, CA, USA, Purdy et al. 1990). The method has been described in details earlier (Timby et al. 2006) and here only briefly described. The antiserum is raised in rabbits against 3 α -hydroxy-20-oxo-5 α -pregnan-11-yl carboxymethyl ether coupled to bovine serum albumin (Purdy et al. 1990). The cross-reactivity of the antibody is shown in earlier publications (Purdy et al. 1990; Timby et al. 2006). The standard curve was established by preparing duplicate tubes containing eight concentrations of unlabeled allopregnanolone to give a range from 0 to 400 pg. Rabbit antiserum was used in a dilution of 1/5,000 and the antibody solutions were prepared with [11,12]³H-allopregnanolone (NEN), 3×10^6 cpm per 32-ml solution containing 65-mM boric acid (Merck) buffer, pH=8.0, bovine serum albumin 100 mg/ml (Sigma), human gamma globulin solution 20 mg/ml (Octapharma), and antibody in milliliter ratio solution of 30:1:1:0.006 making a total solution of 32.006 ml. The rest of the assay was made as described above for isoallopregnanolone. The sensitivity of the assays was 25 pg, with an intra-assay coefficient of variation for allopregnanolone of 6.5% and an interassay coefficient of variation of 8.5%. The recovery of allopregnanolone was 98% and the results are compensated for recovery.

Pharmacokinetic analysis

Baseline serum concentrations (C_0), concentrations 5 min after each intravenous injection (C_5 , C_{35} , and C_{65}), maximum serum concentrations (C_{max}), and the time to achieve maximum serum concentrations (t_{max}) were obtained directly from the measured values. Before the pharmacokinetic calculations were carried out, the baseline isoallopregnanolone and allopregnanolone concentrations were subtracted from the measured values obtained 5–1,860 min later, and only the net concentrations were used for further pharmacokinetic analyses. In all subjects, the baseline isoallopregnanolone concentrations were below 1.9% of the maximum concentrations and the baseline allopregnanolone concentrations were below 1.3% of the maximum concentrations. Pharmacokinetic parameters were calculated by means of the Kinetica program package, version 4.3 (InnaPhase Corporation, Philadelphia, PA, USA), using a noncompartment model. The parameter estimates describing the elimination phase of the log-concentrations of isoallopregnanolone and allopregnanolone (λ_z) were calculated using the concentrations from 330 to 1,860 min after the first dosage, as this gave the best-fit regression lines. The elimination half-lives ($t_{1/2}$) were calculated as $\ln 2/\lambda_z$. Areas under the curve (AUC) were calculated using a mixed log-linear method with extrapolation to infinity. The mean extrapolated AUC was 11.2% of the total AUC for isoallopregnanolone and 22.7% of the

total AUC for allopregnanolone. Clearances (CL) were calculated as dose/AUC and the volumes of distribution (V_d) were calculated as CL/λ_z .

Statistics

Saccadic eye velocity parameters and self-rating scores were calculated as difference from baseline at each time point, e.g., delta degrees per second and delta sedation scores. The daily symptom rating score data were centered on the onset of menstrual bleeding and the test day. The saccadic eye velocity parameters were analyzed by one-way analysis of variance with repeated measures; least significant difference test was used as post hoc test. The SPSS version 11 statistical package was used for analyses. P values of less than 0.05 were considered to be statistically significant.

Results

Seven women were included but only six completed the study. The seventh subject was excluded on the test day because she developed a vasovagal reaction when the intravenous cannula was inserted; she was never given any test injection. Demographic data of the study group are presented in Table 1.

The concentrations of isoallopregnanolone at baseline and after each injection are shown in Table 2 and Figs. 1 and 2, bottom panel. In parallel to increasing concentrations of isoallopregnanolone, there was also a rise in allopregnanolone concentrations with a significant correlation between the isoallopregnanolone and allopregnanolone concentrations ($r=0.484$; $p<0.001$).

The pharmacokinetic parameters of isoallopregnanolone and allopregnanolone are described in Table 2. The first dose of isoallopregnanolone was 0.04 mg/kg, the second dose 0.06 mg/kg, and the third dose 0.10 mg/kg. Five minutes after the first dose, the concentration of isoallo-

Table 1 Demographic data and physical characteristics of the study group ($n=6$)

Age, years, median (range)	30.5 (23–39)
Body mass index, kg/m ² , mean±SEM	24.6±1.7
Married (%)	50
Users of hormone contraceptives (%)	0
Tobacco users (%)	0
Number of children, n , mean±SEM	0.83±0.4
Reported menstrual cycle length, days, mean±SEM	28.8±0.6
Cycle length in test cycle, days, mean±SEM	27.7±1.1
Cycle day when the test was performed, median (range)	9.5 (6–13)

Table 2 Pharmacokinetic parameters of isoallopregnanolone and allopregnanolone in six subjects after three intravenous injections of isoallopregnanolone, representing a cumulative dose of 0.20 mg per kilogram body weight

	Isoallopregnanolone Mean±SD	Allopregnanolone Mean±SD
C_0 (nmol l ⁻¹)	1.01±0.30	0.80±0.13
C_5 (nmol l ⁻¹)	27.5±6.7	3.7±2.1
C_{35} (nmol l ⁻¹)	60.8±17.7	6.6±4.7
C_{65} (nmol l ⁻¹)	138±71.7	19.0±18.4
C_{max} (nmol l ⁻¹)	143±73.0	22.8±22.5
t_{max} (min)	67.7±4.1	74.0±21.9
$t_{1/2}$ (min)	847±599	1,079±527
AUC (nmol l ⁻¹ min)	21,009±6,031	5,853±4,868
CL (ml min ⁻¹ kg ⁻¹)	32.1±9.3	–
V_d (l kg ⁻¹)	36.3±18.1	–

C_0 endogenous serum concentration, C_5 , C_{35} , C_{65} serum concentrations 5 min after each isoallopregnanolone injection, C_{max} maximum serum concentration, t_{max} time to achieve maximum serum concentrations, $t_{1/2}$ elimination half-life, AUC area under the serum concentration–time curve from zero to infinity, CL clearance, V_d volume of distribution

pregnanolone was 27.5 nmol/l; after the second injection, the concentration rose by 33.3 nmol/l and after the third by another 77.2 nmol/l. The theoretic cumulative concentration after the second injection is 68.75 nmol/l (1.5 times the value after the first injection of isoallopregnanolone) and 137.5 nmol/l (2.5 times) at the third injection. These values are very close to the values measured at 5 min after these injections (Table 2), indicating a dose-related increase in the serum concentration of isoallopregnanolone. The maximum serum concentration of isoallopregnanolone was 143 nmol/l, achieved at 67.7 min. The maximum serum concentration of

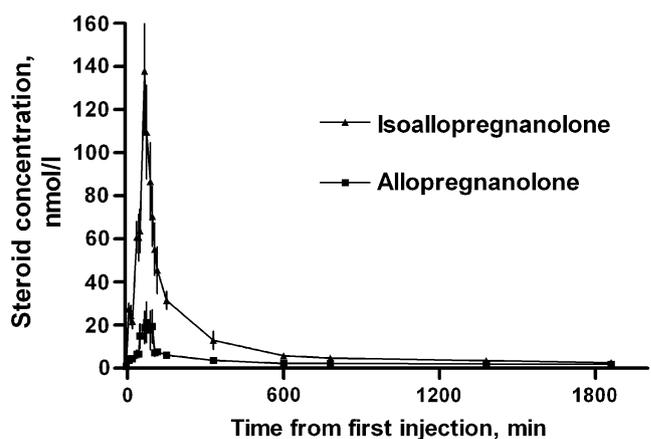


Fig. 1 Mean serum concentrations of isoallopregnanolone (nmol/l) and allopregnanolone (nmol/l) measured at 19 time points. Concentrations are measured at baseline (–5 min) before the first injection (0 min) and until 1,860 min after the first injection. Maximum concentration occurred at 67.7±4.1 min for isoallopregnanolone and at 74.0±21.9 min for allopregnanolone

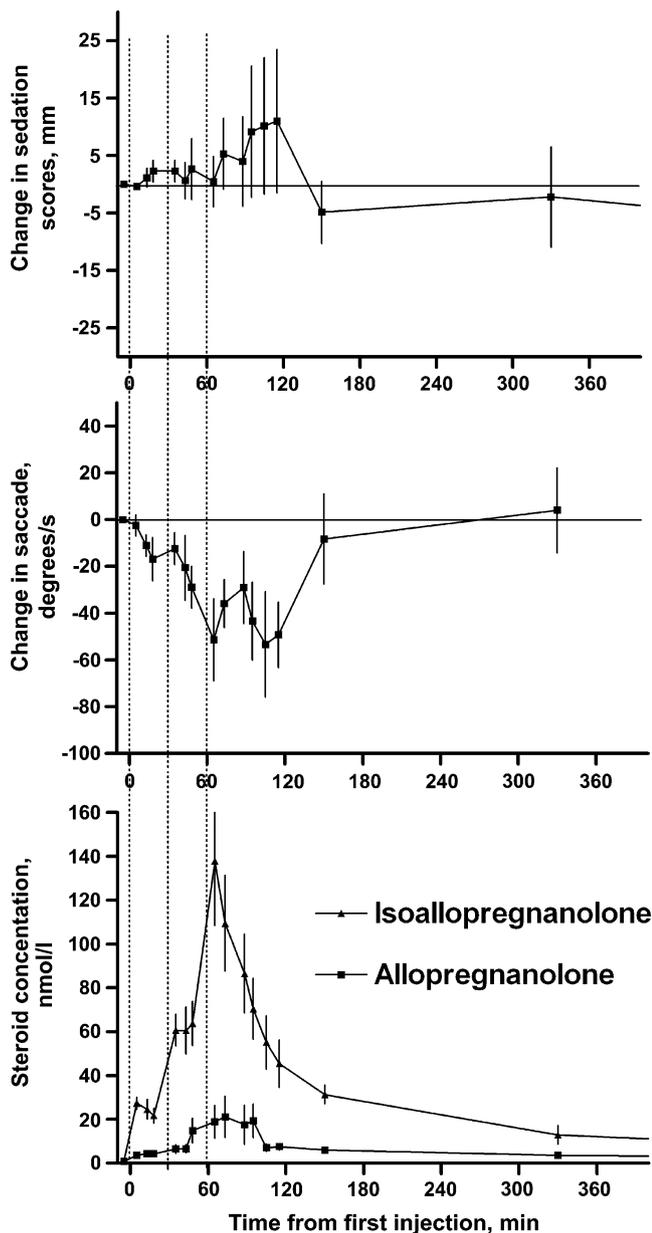


Fig. 2 Mean (SEM) effects of isoallopregnanolone injections in women in the follicular phase on scored sedation (*top panel*) and saccadic eye velocity (*middle panel*) compared to the steroid concentrations (*bottom panel*). Dose 1 (0.04 mg/kg), dose 2 (0.06 mg/kg), and dose 3 (0.10 mg/kg) were given at 0, 30, and 60 min and are indicated in the figure as *dotted lines*. No significant change in scored sedation was noted. There was an overall decrease in SEV from 5 to 115 min after the first injection ($F(14, 70)=3.27$; $p<0.001$)

allopregnanolone was 22.8 nmol/l, which is 16% of the maximum concentration of isoallopregnanolone, and occurred 6 min later, at 74.0 min.

In Fig. 2, top panel, the effects on sedation are shown. No significant changes occurred in sedation following the three injections of isoallopregnanolone.

Figure 2, middle panel, shows the changes in saccadic eye velocity during the test. There was an overall significant decrease in SEV ($F(14, 70)=3.27$; $p<0.001$). The post hoc test showed significant decreases in SEV 5 and 13 min after the first injection ($p=0.014$ – 0.027) and 5 and 18 min after the second injection ($p=0.007$ – 0.024). After the third injection, there were significant decreases during the first 35 min ($p=0.019$ – 0.049), not at 45 min, but then again at 55 min ($p=0.008$). From 150 min after the first injection, SEV had returned to baseline again.

The effect on vital functions is shown in Table 3. There were no significant changes in the systolic and diastolic blood pressure after the injections, although there was a significant decrease in heart rate during the course of the study day (Table 3, $F(5, 25)=6.12$; $p<0.001$). The respiration frequency showed no significant change. The menstrual cycle length of the test cycle was 27.7 ± 1.1 days, which was not significantly different from the reported cycle length of 28.8 ± 0.6 days prior to the test (Table 1, mean \pm SEM).

During the test, the six women performed four different psychological tests. With the ASI test at baseline, all women were within a low anxiety range (score 0–10), except one subject with a score of 12. The ASI test has a total range from 0 to 64 (Grant et al. 2007). PSS and STAI revealed no significant changes from baseline until the day after the test (PSS 0.33 ± 0.33 versus 0.33 ± 0.21 ; STAI 41.8 ± 2.1 versus 42.8 ± 2.6). No fluctuations in anxiety level were revealed by SADS when used repeatedly from baseline through the test day and until 31 h after the first injection.

To investigate whether isoallopregnanolone had any effect on mood symptoms, the data were centered around the day of the test injection. Summarized positive, negative,

Table 3 Vital functions (mean \pm SEM) before and at several occasions after intravenous injections of isoallopregnanolone in six women during the follicular phase

Occasion	Systolic blood pressure, mmHg	Diastolic blood pressure, mmHg	Heart rate, beats/min	Respiration rate, number/min
Baseline	103 \pm 6.5	73 \pm 3.5	67 \pm 4.3 ^a	19 \pm 1.3
5 min after first inj	107 \pm 5.1	72 \pm 3.6	65 \pm 3.7	15 \pm 0.9
5 min after second inj	103 \pm 3.5	71 \pm 2.2	61 \pm 4.0	14 \pm 1.0
5 min after third inj	102 \pm 3.9	71 \pm 3.0	57 \pm 4.1	17 \pm 1.9
35 min after third inj	102 \pm 3.9	70 \pm 3.0	57 \pm 3.5	16 \pm 1.0
9 h after third inj	106 \pm 2.6	69 \pm 4.1	68 \pm 3.3	13 \pm 1.5

^a Statistically significant ($F(5, 25)=6.12$; $p<0.001$) change in heart rate during the course of the study day

and physical symptoms as well as the individual symptoms were investigated from 5 days prior to the test until 10 days after the test are shown in Fig. 3. There were no significant changes in any symptoms after the injections.

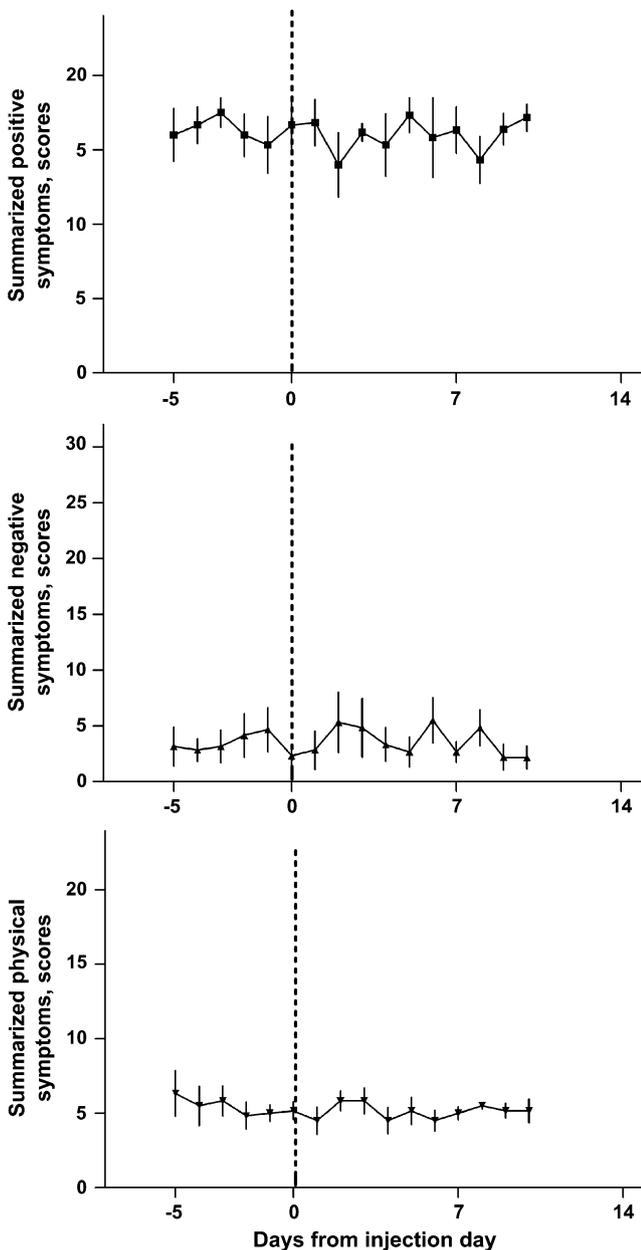


Fig. 3 Mean (SEM) summarized positive, negative, and physical symptoms from the daily ratings are plotted and centered round the test day in the follicular phase, 5 days prior to the test and 10 days after. There were no significant changes after the injections compared to the pretest scores. Daily ratings were made for menstrual bleeding and 15 other physical and mood changes during the whole menstrual cycle in which the experiment was performed. Scores for cheerfulness, happiness, energy, and interest in daily activities are summarized and plotted as positive symptoms (0–32). Scores for irritability, tension, fatigue, and depressed mood are summarized and plotted as negative symptoms (0–32). Breast tenderness, change in appetite, and swelling are summarized and plotted as physical symptoms (0–24)

Summarized positive, negative, and physical symptoms were plotted in an ideal menstrual cycle of 28 days by centering the score data around the onset of menstruation (data not shown). Summarized positive symptoms showed no significant variation during the cycle. Summarized negative mood and physical symptoms showed a slight but significant overall variation within the cycle ($F(27, 135)=1.62$; $p=0.039$ and $F(27, 135)=2.34$; $p<0.001$, respectively). For summarized negative mood symptoms, there was a significant overall variation but no single day showed significant symptom severity over the other days. For summarized physical symptoms, the first and the last day of the cycle showed a significant symptom increase compared to the follicular phase ($p<0.05$, $p=0.01$).

When statistics were calculated for each of the 16 symptoms except menstrual bleeding, in the ideal menstrual cycle, some of the symptoms showed significant variation. These were breast tenderness ($F(27, 135)=0.300$; $p<0.001$), swelling ($F(27, 135)=2.63$; $p<0.001$), and sleeping problems ($F(27, 135)=1.66$; $p=0.032$). Decreased interest for daily activities ($F(27, 135)=2.32$; $p=0.001$) was only significant at onset of the menstrual bleeding. Daily life impairment showed significant variation ($F(27, 135)=2.46$; $p<0.001$) being higher in cycle days 1 and 3 ($p=0.046$ – 0.022 and $p=0.042$ – 0.041) compared with the last 2 days of the cycle. Cycle days 27 and 28 had significant less impairment then cycle days 1–3.

Discussion

The pharmacokinetics of isoallopregnanolone, given as an intravenous injection to humans, has not been studied before. One of the main findings of the present study was a dose-proportional increase in the serum concentration of isoallopregnanolone after intravenous administration of a cumulative dose of 0.20 mg/kg. A second finding was a rather slow elimination of isoallopregnanolone, with a half-life of about 850 min. There was a difference in response to isoallopregnanolone on saccadic eye velocity compared to self-rated sedation. No changes related to isoallopregnanolone injection were noted in the mood symptom ratings.

In parallel to the rise in concentration of isoallopregnanolone, but with a slight delay, there was also a rise in serum concentration of allopregnanolone. The obvious explanation is that some isoallopregnanolone has been metabolized to allopregnanolone. This theory is supported by earlier findings showing that allopregnanolone and isoallopregnanolone serum concentrations are highly correlated both during the follicular and the luteal phase of the menstrual cycle (Havlikova et al. 2006). Furthermore, the 3- α hydroxylase is effective in both direction and when allopregnanolone is given to animals there is an increase in

isoallopregnanolone concentration so one can expect that when isallopregnanolone is given allopregnanolone should increase (Vallee et al. 2000).

In the present study, clearance of isallopregnanolone was 32.1 ml/min per kilogram. This result should be compared with previously reported values of 32.6 ml/min per kilogram for allopregnanolone (Timby et al. 2006) and 61.0 ml/min per kilogram for pregnanolone (Sundstrom et al. 1999b). Thus, the clearance of allopregnanolone and isallopregnanolone appears to be about the same, indicating a similar elimination rate of the two substances.

Allopregnanolone has a hydroxyl group in a planar 3α position compared to the angle position (the 3β position) in relation to the steroid skeleton for isallopregnanolone. Steroids with a hydroxy group in the 3α position have been reported to be glucuronized at a higher degree than steroids with a hydroxy group in the 3β position (Jin et al. 1997), most likely because the 3α position better fits the active site on the conjugation enzyme. The amount of allopregnanolone glucuronized might therefore be higher than the corresponding amount of isallopregnanolone. As the clearance values for these two substances nevertheless are about the same, other metabolic pathways (or active renal excretion pathways) would be expected to contribute to the elimination of isallopregnanolone to a higher degree than for allopregnanolone. Such a metabolic pathway might be degradation via cytochrome P-450 3A4 (CYP3A4), which is known to be involved in the metabolism of many sex hormones (Yamazaki and Shimada 1997; Niwa et al. 1998; Tsuchiya et al. 2005).

The elimination half-life of allopregnanolone in the present study was about 1,100 min, which is more than four times the elimination half-life of allopregnanolone of 260 min reported previously (Timby et al. 2006). This most obvious reason is a continuous production of allopregnanolone from isallopregnanolone in the present study, implying that the half-life calculated for allopregnanolone is not based upon data from a true elimination phase.

Compared to the previously reported elimination half-life of allopregnanolone (Timby et al. 2006), the elimination half-life of isallopregnanolone of about 850 min in the present study is also rather long. The solvent might be of importance for the half-life by creating a possible depot from which the substance is slowly released, as shown for pregnanolone (Carl et al. 1994; Sundstrom et al. 1999b). However, both allopregnanolone and isallopregnanolone were administered in a similar albumin solution. Thus, the most likely explanation for the differences in elimination half-lives, despite the same clearance values, is a higher volume of distribution for isallopregnanolone. In fact, the volume of distribution for isallopregnanolone in the present study was 36 l/kg, whereas it

was 12.5 l/kg for allopregnanolone in the study by Timby et al. (2006). We have not investigated the tissue distribution in more detail in the present study, but the differences in retention times in the HPLC suggest somewhat higher lipid solubility for isallopregnanolone.

The method used for the isallopregnanolone assay is RIA after HPLC separation of cross-reacting steroids. The antiserum used is made against pregnenolone and therefore not specific against isallopregnanolone but binds isallopregnanolone sufficiently for a standard curve to be obtained. To avoid overestimation of isallopregnanolone by pregnenolone and cross-reacting steroids, a HPLC separation was used in advance. A number of cross-reacting steroids were separated from isallopregnanolone with this procedure (see “Materials and methods”). The study was carried out in the follicular phase of the menstrual cycle when the endogenous concentrations of isallopregnanolone and allopregnanolone are low. The baseline endogenous concentration obtained in the present study is in the upper range of concentrations reported in the literature when using HPLC–RIA and gas chromatography–mass spectrometry as analytical techniques (Romeo et al. 1998; di Michele et al. 2003; Schule et al. 2003; Murphy et al. 2004; Havlikova et al. 2006). The concentrations measured in the present study are also higher than those obtained during the mid follicular phase in women at noon using the same assay method as in the present study (0.22 ± 0.77 nmol/l (mean \pm SD), range 0.1 to 1.1 nmol/l, $n=77$, unpublished results). In addition, the allopregnanolone concentrations (Table 2) are in the upper normal range at baseline and higher than levels during mid follicular phase in women but in the same range as samples taken in early follicular phase (Nyberg et al. 2007).

There are two factors that may explain the baseline levels. The first factor is that the baseline samples are all taken early in the morning, that is, at a time when adrenocorticotrophic hormone (ACTH)-stimulated steroid production from the adrenals is high. During the follicular phase, the ACTH-stimulated adrenals are the major source of serum allopregnanolone and most likely also of isallopregnanolone (Torres et al. 2001; Genazzani et al. 2002). The second factor is that the situation was somewhat stressful. The subjects knew that they were about to receive a compound never given to humans before, and electrodes for oculography had been mounted and intravenous cannulas inserted prior to testing. It is known that allopregnanolone and most likely also isallopregnanolone increase after stress both in humans and animals (Droogleever et al. 2004; Purdy et al. 1991). The allopregnanolone concentrations at baseline in our subjects are slightly lower than in amenorrhic patients known to have a high adrenal stimulation (Genazzani et al. 2002). These factors may explain the somewhat higher baseline

concentrations of isoallopregnanolone and allopregnanolone in the follicular phase in the present study.

However, after the injections exogenously administered, isoallopregnanolone will dominate in the sample and the most likely metabolites and cross-reacting steroids like pregnenolone and allopregnanolone were separated with the preceding HPLC.

The current study shows no significant influence on self-rated sedation after injections of isoallopregnanolone, but there is a significant decrease in saccade velocity during the test. The effect on SEV was not anticipated, as earlier *in vitro* studies by our group have not shown any effect, whatsoever, directly on the GABA_A receptor (Lundgren et al. 2003). A possible explanation to this finding is that metabolism of isoallopregnanolone to allopregnanolone occurs and that the effect on the saccade velocity actually is an effect of allopregnanolone. This theory is supported by the course of events during the study. The effect on SEV lasted from the first injection of isoallopregnanolone until 150 min after the last injection, which is longer than the effect produced by allopregnanolone directly where the effect declined already after 45 min (Timby et al. 2006). In the present study, as well as in the Timby study, the maximum concentrations of the steroid injected were reached 5 min after the last injection. In the Timby study, the maximum concentration of allopregnanolone was about 70 nmol/l after a cumulative dose of 0.09 mg/kg and the maximum concentration of isoallopregnanolone in our study was almost 140 nmol/l after a cumulative dose of 0.20 mg/kg. However, in the present study, there was a delay of 6 min for the concentration peak of allopregnanolone (20 nmol/l) to occur compared to the isoallopregnanolone peak. The concentration of isoallopregnanolone decreased quite fast, but for allopregnanolone the concentration made a plateau and was almost constant until 35 min after the last isoallopregnanolone injection (Fig. 2 bottom panel). In addition, the cumulative dose of isoallopregnanolone was quite high, which might facilitate a substantial metabolism to allopregnanolone. Finally, an additional metabolism of isoallopregnanolone to allopregnanolone might also take place within the central nervous system without being detected in the periphery. The 3- α -hydroxylase is effective in both directions, and the conversion of isoallopregnanolone to allopregnanolone is probably higher in the brain than in plasma. A higher conversion of allopregnanolone to isoallopregnanolone in the brain compared to plasma has been shown in rodents and it is likely that a similar conversion exists in humans (Vallee et al. 2000). Due to high concentration of isoallopregnanolone, the metabolism is most probably in the direction from isoallopregnanolone to allopregnanolone and a higher allopregnanolone concentration can be assumed in the brain than in plasma. However, we cannot exclude the possibility that isoallopregnanolone

has an effect of its own, even though earlier *in vivo* and *in vitro* studies have not been able to show it (Gyermek 1968; Lundgren et al. 2003; Stromberg et al. 2006).

Interestingly, no effect on sedation was noted in the present study, although a significant decrease in SEV was observed. Several explanations to this finding are possible. The lack of effect on sedation might be due to an antagonistic effect of isoallopregnanolone on allopregnanolone. *In vivo* studies in rats show an antagonistic effect of isoallopregnanolone on allopregnanolone-induced anesthesia (Backstrom et al. 2005). Another explanation to the lack of effect on sedation is that the concentration of allopregnanolone was less than one third of the concentrations produced in earlier studies where a distinct effect on sedation was found (Timby et al. 2006). Thus, higher concentrations than in the present study might be necessary for such an effect. SEV might also be a more sensitive measurement of sedation than subjective VAS scores.

It has previously been shown that sedation and saccadic eye velocity can show different responses to GABA_A stimulation. For instance, thyrotropin-releasing hormone reverses benzodiazepine-induced sedation but not the slowing of saccadic eye velocity (Glue et al. 1992). An explanation to the differential effects on sedation and SEV might be differences in metabolism and/or receptor interaction in different areas of the brain so that the antagonistic effect is more pronounced in areas involved in sedation compared to areas controlling the saccadic eye velocity.

Since one of our hypotheses is that allopregnanolone, isoallopregnanolone, and other neurosteroids are involved in the etiology of PMDD, it was important to detect any effect on mood and anxiety during the experiment. However, no acute effects were noted and no delayed effects were seen during a period of 10 days after the experiment. It is also interesting to notice that there was no significant change in the length of the menstrual cycle or in mood and physical symptoms during the menstrual cycle in which the test was performed. Even though some cyclic symptom changes were observed at the end of the menstrual cycle, these were considered to be the normal cyclic symptom variations seen in most women premenstrually. No significant changes occurred on the test day or during the days following the test.

Isoallopregnanolone seems to be safe when given intravenously at a cumulative dose of 0.20 mg/kg, as no significant adverse events were reported and no significant effects on vital functions were noted. A slight decrease in heart rate was detected after each injection but that was probably an effect of decreased stress and rest during the test day. However, it should be emphasized that the number of subjects included is far too low to detect infrequent

adverse reactions. Moreover, if given in higher doses, the frequency of adverse reactions might increase.

In conclusion, isoallopregnanolone seems to be safe to give intravenously to healthy women at the present doses and does not interfere with the menstrual cycle. For the first time, pharmacokinetic properties of isoallopregnanolone have been investigated in humans. Finally, isoallopregnanolone has an effect on saccadic eye movement, although not on self-rated sedation, most probably due to the metabolism to allopregnanolone.

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