

# Acute Effects of Riluzole and Retigabine on Axonal Excitability in Patients With Amyotrophic Lateral Sclerosis: A Randomized, Double-Blind, Placebo-Controlled, Crossover Trial

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Increased excitability of motor neurons in patients with amyotrophic lateral sclerosis (ALS) may be a relevant factor leading to motor neuron damage. This randomized, double-blind, three-way crossover, placebo-controlled study evaluated peripheral motor nerve excitability testing as a biomarker of hyperexcitability and assessed the effects of riluzole and retigabine in 18 patients with ALS. We performed excitability testing at baseline, and twice after participants had received a single dose of either 100 mg riluzole, 300 mg retigabine, or placebo. Between- and within-day repeatability was at least acceptable for 14 out of 18 recorded excitability variables. No effects of riluzole on excitability testing were observed, but retigabine significantly decreased strength-duration time-constant (9.2%) and refractoriness at 2 ms (10.2) compared to placebo. Excitability testing was shown to be a reliable biomarker in patients with ALS, and the acute reversal of previously abnormal variables by retigabine justifies long-term studies evaluating the impact on disease progression and survival.

## Study Highlights

### WHAT IS THE CURRENT KNOWLEDGE ON THE TOPIC?

☑ Patients with amyotrophic lateral sclerosis show increased membrane excitability in peripheral and central motor neurons, which may be a relevant factor leading to motor neuron damage. Current approved treatment with riluzole decreases hyperexcitability, indicating this could be a therapeutic target.

### WHAT QUESTION DID THIS STUDY ADDRESS?

☑ The aim of this study was to validate peripheral motor nerve excitability testing as biomarker of hyperexcitability and assess effects of riluzole and retigabine in patients with amyotrophic lateral sclerosis.

### WHAT DOES THIS STUDY ADD TO OUR KNOWLEDGE?

☑ Between- and within-day repeatability was good for most excitability testing parameters and a single dose of retigabine, but not riluzole normalized several relevant markers indicating hyperexcitability in these patients.

### HOW MIGHT THIS CHANGE CLINICAL PHARMACOLOGY OR TRANSLATIONAL SCIENCE?

☑ Peripheral nerve excitability could be a very useful and quick, noninvasive biomarker to test for potential treatments for ALS, and measure treatment efficacy. Retigabine may be such a treatment reversing hyperexcitability, justifying long-term studies looking at the impact on disease progression and survival.

Amyotrophic lateral sclerosis (ALS) is a progressive and fatal neurodegenerative disease characterized by ongoing loss of motor neurons. In ALS the unique phenomenon of increased membrane excitability in both peripheral and central motor neurons can be observed, presenting clinically as fasciculations, muscle cramps, hyper-reflexia, and spasticity.<sup>1</sup> Excitability testing is a neurophysiological tool that allows noninvasive assessment of axolemmal voltage-gated ion-channel activity in motor axons of a peripheral nerve. In ALS, it showed evidence of increased persistent sodium-conductance and reduced potassium-conductance,

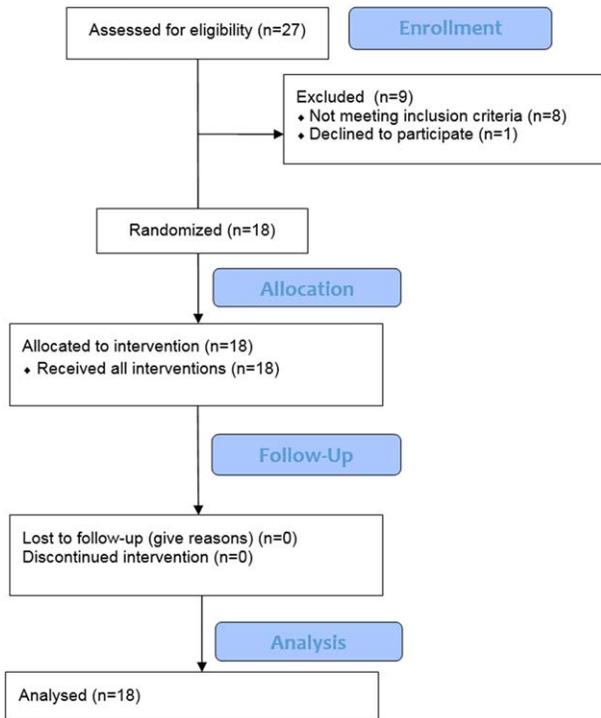
both of which may contribute to axonal hyperexcitability and fasciculation.<sup>2–6</sup> Furthermore, the presence of increased persistent sodium-conductance was shown to be correlated with more rapid functional decline and shorter survival,<sup>7,8</sup> and the presence of fasciculation with shorter survival.<sup>9</sup> Because increased membrane excitability in ALS may be a relevant step in the cascade leading to structural damage of motor neurons,<sup>10</sup> early identification of hyperexcitable motor neurons may provide an argument for initiating neuroprotective intervention.<sup>7,11</sup> Retigabine, a potassium-channel activator, was shown to reduce increased cellular

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**Figure 1** Study flow diagram. [Color figure can be viewed at [cpt-journal.com](http://cpt-journal.com)]

membrane excitability and improve cell survival in an *in vitro* model of ALS.<sup>12</sup> Riluzole, the only registered drug for the treatment of ALS, partially normalized some excitability variables in peripheral and cortical motor neurons of patients with ALS.<sup>13</sup> Modulation of hyperexcitability could therefore serve as a proof-of-pharmacology biomarker to assess the effects of therapeutic interventions in ALS. The present study aimed to validate motor nerve excitability testing of the median nerve as a biomarker of hyperexcitability and assessed the pharmacodynamic effects of retigabine and riluzole in patients with ALS.

## RESULTS

The interim analysis showed at least acceptable repeatability ( $\alpha > 0.7$ ) for all five predetermined variables (data not shown). Therefore, in total, 18 patients with ALS were included and all subjects completed the study (**Figure 1**), with recruitment running from October 2015 to December 2016, and the last follow-up phone call taking place in April 2017. Baseline characteristics are displayed in **Table 1**. Participants tolerated the study and treatments well. One subject did not complete the 6-hour measurement of the first visit (riluzole occasion) because of adverse events consisting of nausea and vomiting due to a migraine attack. It was considered unlikely that this was related to the study treatment. There were 15 adverse events in the retigabine arm, 14 in the riluzole arm, and six in the placebo arm. All events were grade 1–2 and none were reported more than twice per arm, except for dizziness (reported three times in the retigabine arm) and somnolence (reported seven times in the retigabine and three times in the placebo arm).

**Table 1** Demographics

N	18
Age (years)	58.6 (37–76)
Weight (kg)	85.0 (8.2)
Height (cm)	182.1 (6.7)
Sex (Female/Male)	1 (6%) / 17 (94%)
Time since symptom onset (months)	28.9 (6.9–106.7)
Time since diagnosis (months)	13.7 (3.0–62.5)
Taking standard riluzole treatment	17 (94%)
Riluzole treatment duration (months)	12.8 (2–61)
ALSFRS–R at baseline	38 (29–45)
Familial history of ALS (Yes/No)	4 (22%) / 14 (78%)

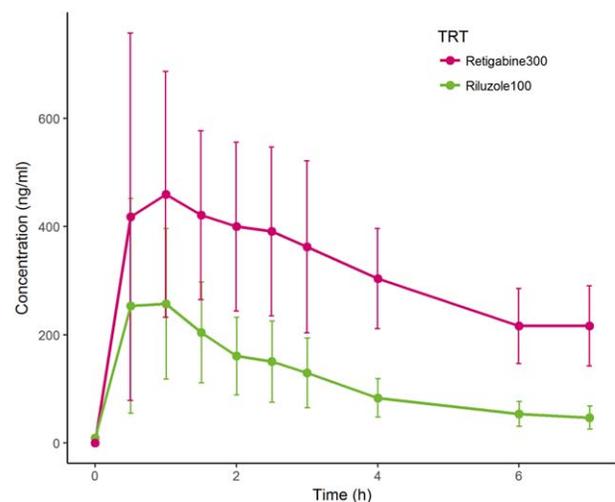
Values are presented as mean (SD or range where appropriate).

## Pharmacokinetics

Pharmacokinetic analysis showed mean maximum plasma concentration ( $C_{max}$ ) for riluzole of 343 ng/mL (range: 102–646 ng/mL) and for retigabine of 604 ng/mL (271–997 ng/mL), both on average at 1 hour postdose (riluzole range: 0.5–3 hours; retigabine range: 0.5–4 hours) (**Figure 2**). Variability in  $C_{max}$  was moderate, with a coefficient of variation (CoV) of 48% for riluzole and 36% for retigabine. Time to  $C_{max}$  ( $T_{max}$ ) was more variable for retigabine (CoV 90%) than for riluzole (CoV 66%). Mean riluzole concentration before dosing for all treatment periods combined was 12 ng/mL (range: <1–75.7 ng/mL), and 10 ng/mL (1.9–22.3 ng/mL) for the placebo treatment period.

## Baseline electrophysiological characteristics and test–retest reliability

Baseline electrophysiological characteristics, as well as repeatability for excitability testing are presented in **Table 2**. Repeatability between each baseline measurement (interoccasion), as



**Figure 2** Pharmacokinetics of riluzole and retigabine. Mean with standard deviation concentration–time profiles in plasma per treatment (TRT).

**Table 2 Baseline excitability characteristics, variability, and repeatability**

Parameter	Mean	Intersubject CV (%)	Intrasubject CV (%)	Model-based intrasubject CV (%)	Interoccasion Cronbach's alpha	Intraoccasion Cronbach's alpha
CMAP (mV)	7.2	54.2	25.8	13.9 <sup>a</sup>	0.96	0.97
Threshold for 50% CMAP (mA)	5.95	52.7	39.2	21.8 <sup>a</sup>	0.68	0.90
<i>Strength-duration</i>						
Rheobase (mA)	3.98	57.6	43.7	23.3 <sup>a</sup>	0.68	0.89
SDTC (ms)	0.459	14.8	11.0	8.3 <sup>a</sup>	0.69	0.67
<i>Threshold electrotonus</i>						
TEdpeak (%)	68.8	8.8	4.7	3.8	0.90	0.94
S2 accommodation (%)	20.8	20.4	14.1	12.4	0.80	0.84
Accommodation half-time (ms)	38.4	12.9	11.8	10.5	0.36	0.43
TEd40-60 (%)	52.5	9.6	6.0	4.6	0.84	0.90
TEd90-100 (%)	47.9	11.6	6.0	6.1	0.91	0.95
TEh90-100 (%)	-127.9	21.3	8.7	7.5	0.94	0.95
Fanning (%)	171.6	16.3	7.8	5.7	0.93	0.95
<i>Current-threshold relation</i>						
Resting I/V-slope	0.55	20.3	11.5	7.3 <sup>a</sup>	0.81	0.92
Minimum I/V-slope	0.24	27.3	12.3	10.3 <sup>a</sup>	0.92	0.93
Hyperpolarizing I/V-slope	0.33	27.2	19.0	20.0 <sup>a</sup>	0.79	0.89
<i>Recovery cycle</i>						
Refractoriness at 2 ms (%)	41.9	62.5	45.5	20.9	0.73	0.83
Superexcitability (%)	-26.4	29.7	11.9	9.5	0.95	0.94
Subexcitability (%)	11.7	36.9	22.3	18.5	0.84	0.84
Refractory period (ms)	2.6	11.9	7.7	4.7	0.81	0.91

Excitability variables mean and inter- and intrasubject CV based on baseline measurements at each visit, and intrasubject variability based on the statistical model. Cronbach's alpha for each excitability parameter calculated for each of the three baseline measurements (interoccasion) and for the three measurements within the placebo visit (intraoccasion). Repeatability based on Cronbach's alpha: < 0.5, unacceptable; 0.5–0.6, poor; 0.6–0.7, questionable; 0.7–0.8, acceptable; 0.8–0.9, good; 0.9–1.0 excellent.

<sup>a</sup>Intrasubject variability of the LOG-transformed data.

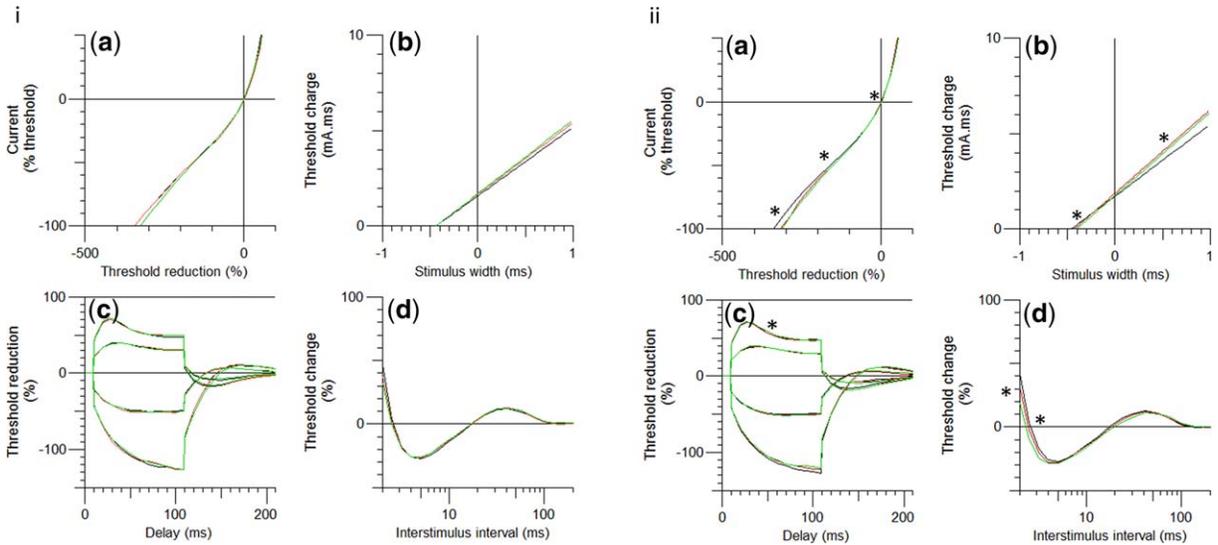
determined by Cronbach's alpha, showed that the majority of variables had an acceptable (Cronbach's alpha >0.7) to excellent repeatability. Only accommodation half-time showed Cronbach's alpha much lower than 0.7; rheobase, strength-duration time constant (SDTC), and threshold for a target Compound muscle action potential (CMAP) of 50% were just below the threshold of Cronbach's alpha >0.7. Within the placebo visit (intraoccasion), repeatability was just below acceptable for SDTC, and well below 0.7 for accommodation half-time, but good to excellent for all other parameters. When compared to controls, the excitability variables obtained in ALS patients at the predose visit showed greater changes in TEd40-60 ( $53.0 \pm (\text{SEM}) 1.2\%$  vs.  $47.3 \pm 0.9\%$ ,  $P < 0.001$ ) and TEd90-100 ( $48.8 \pm 1.6\%$  vs.  $44.1 \pm 0.7\%$ ,  $P = 0.008$ ), longer accommodation half-time ( $38.7 \pm 1.0$  ms, vs.  $35.8 \pm 0.7$  ms,  $P = 0.02$ ), increased superexcitability ( $-27.4 \pm 2.1\%$ , vs.  $20.5 \pm 1.0\%$ ,  $P = 0.004$ ), and decreased subexcitability ( $11.9 \pm 1.0\%$  vs.  $16.7 \pm 1.3\%$ ,  $P = 0.006$ ). Other variables, including SDTC ( $0.46 \pm 0.02$  ms vs.  $0.45 \pm 0.01$  ms,  $P = 0.86$ ), were not significantly different.

### Effects of riluzole and retigabine on motor nerve excitability

**Figure 3i** shows the mean excitability recordings at predose and after a single dose of 100 mg riluzole at 1.5 and 6 hours. No statistically significant effects were observed for riluzole on any of the excitability measures compared to placebo. **Figure 3ii** shows the mean excitability recordings at predose and after a single dose of 300 mg retigabine at 1.5 and 6 hours. Significant treatment effects were observed for retigabine, showing the following effects compared to placebo: increase in hyperpolarizing I/V-slope (21.7%), resting I/V-slope (6.1%), minimum I/V-slope (8.5%), rheobase (28.0%), threshold for a target CMAP of 50% (25.0%), accommodation half-time (3.15 ms), decrease in SDTC (9.2%), refractoriness at 2 ms (10.2% (arithmetic difference)) and refractory period (0.17 ms) (**Table 3, Figure 4**).

### Predictive value of excitability measures

None of the excitability variables at baseline showed a significant correlation with clinical deterioration by a functional decline in revised ALS functional rating scale (ALSFRS-R) score between baseline and 3 months.



**Figure 3** Mean excitability recordings predose (black), and 1.5 hours (red) and 6 hours (green) after a single dose of 100 mg riluzole (i) and 300 mg retigabine (ii). (a) current/voltage relationship, (b) strength-duration properties plotted as stimulus charge vs. stimulus duration, (c) threshold electrotonus, (d) recovery cycle. Asterisks indicate significant treatment effects on thresholds (see **Table 3**).

## DISCUSSION

Excitability testing was shown to produce repeatable results in patients with ALS, both within and between visits, for all 18 variables except accommodation half-time. Although riluzole did not show effects, retigabine had significant effects on several excitability variables when compared to placebo. No correlation between ALSFRS-R and excitability variables was found.

### Electrophysiological characteristics and variability

Intersubject variability for parameters such as CMAP, refractoriness at 2 ms, rheobase, I/V-slopes, super- and subexcitability was relatively high (CV of 27–63%), which may likely be related to differences in the disease state of the patients. Intrasubject variability, therefore, is more informative on variability, and CVs were indeed much smaller. The statistical model-based estimate of the intrasubject CV for all parameters ranged from 4–23%, and for most CVs was very similar to the statistical model-based estimate found in healthy volunteers.<sup>14</sup> Only the CVs of accommodation half-time, TE<sub>d90-100</sub>, TE<sub>h90-100</sub>, and hyperpolarizing I/V slope were more than 1.5 times greater than found by Tomlinson *et al.*<sup>14</sup>

### Pharmacokinetics

The  $C_{max}$  of riluzole for all but three subjects (mean 343 ng/mL, range 102–646 ng/mL) and of retigabine for all subjects (mean 604 ng/mL, range 271–997 ng/mL) was above the approximate therapeutic levels of 173 ng/mL for riluzole<sup>15</sup> and 250 ng/mL for retigabine,<sup>16</sup> as expected with the selected supratherapeutic doses. Mean plasma concentration of riluzole before dosing for all treatment periods was 12 ng/mL (range <1–75.7 ng/mL), as could be expected after ~24-hour washout of riluzole; thereby, levels were ~30 times lower than at  $C_{max}$  during the riluzole period. As the riluzole concentration exerting 50% of the maximal effect ( $IC_{50}$ ) on voltage gated sodium channels is ~0.3  $\mu$ M

(or 70 ng/mL),<sup>17</sup> the mean post-washout level of 12 ng/mL is unlikely to have affected sodium channel function. Two subjects had significantly higher riluzole baseline levels at one visit (with 63.2 and 75.7 ng/mL), possibly due to not having followed the instruction to omit their regular evening dose of riluzole. Both instances occurred during retigabine treatment periods. It is, however, unlikely that this influenced the observed effects in the retigabine treatment arm, especially as no effect of riluzole was observed in the riluzole treatment arm. Sensitivity analysis, with exclusion of these occasions, did not produce a different outcome, with one exception: that retigabine effects on minimum I/V-slope and resting I/V-slope did not reach significance (borderline) (data not shown).

### Effects of retigabine on peripheral nerve excitability

In human peripheral motor nerve, five types of voltage-gated potassium channels have been described, depending on their gating modes, activation-deactivation time, and conductance, with a large overlap between their kinetic properties.<sup>18,19</sup> These five types give rise to three types of potassium currents on single axon recordings: fast, intermediate, and slow. Slowly activating potassium channels of the axonal membrane belong to Kv7.2–Kv7.5 subtypes, coded by the KCNQ genes.<sup>20,21</sup> *In vitro* studies and animal models of epilepsy and pain showed that retigabine hyperpolarizes resting axonal membrane potential by inducing these potassium channel subtypes to open which, in turn, enhances outward slow potassium currents and produces a hyperpolarizing shift of the half-activation potential of these channels.<sup>22,23</sup> Excitability variables assessed after potassium channel activators such as retigabine and flupirtine administration, therefore, are expected to reflect either potassium channel activation or the resulting hyperpolarization of resting membrane potential.<sup>20,21</sup>

**Table 3 Treatment effects on parameters of motor nerve excitability**

Parameter	Contrast retigabine vs. placebo	Contrast riluzole vs. placebo
CMAP (mV)	5.64 vs. 5.82 -3.1% (-14.0%, 9.1%) P = 0.584	5.24 vs. 5.82 -9.9% (-20.5%, 2.1%) P = 0.097
Threshold for 50% CMAP (mA)	5.97 vs. 4.78 25.0% (7.6%, 45.2%) P = 0.005	5.54 vs. 4.78 16.1% (-0.3%, 35.2%) P = 0.055
<i>Strength-duration</i>		
Rheobase (mA)	4.02 vs. 3.14 28.0% (9.1%, 50.1%) P = 0.004	3.63 vs. 3.14 15.4% (-2.0%, 35.7%) P = 0.083
SDTC (ms)	0.416 vs. 0.458 -9.2% (-14.1%, -3.9%) P = 0.001	0.455 vs. 0.458 -0.6% (-6.1%, 5.1%) P = 0.821
<i>Threshold electrotonus</i>		
TEdpeak (%)	68.31 vs. 69.42 -1.12 (-3.03, 0.80) P = 0.241	69.09 vs. 69.42 -0.33 (-2.27, 1.61) P = 0.730
S2 accommodation (%)	21.29 vs. 20.52 0.77 (-1.14, 2.68) P = 0.415	19.44 vs. 20.52 -1.08 (-3.02, 0.85) P = 0.260
Accommodation half-time (ms)	39.88 vs. 36.74 3.15 (0.91, 5.38) P = 0.007	36.50 vs. 36.74 -0.24 (-2.51, 2.03) P = 0.831
TEd40-60 (%)	53.29 vs. 53.36 -0.070 (-1.849, 1.709) P = 0.936	52.96 vs. 53.36 -0.399 (-2.200, 1.401) P = 0.652
TEd90-100 (%)	47.09 vs. 48.84 -1.748 (-3.867, 0.372) P = 0.102	49.73 vs. 48.84 0.896 (-1.259, 3.051) P = 0.401
TEh90-100 (%)	-121.60 vs. -122.70 1.093 (-5.722, 7.907) P = 0.746	-127.71 vs. -122.70 -5.016 (-11.872, 1.840) P = 0.146
Fanning (%)	164.16 vs. 168.45 -4.28 (-11.85, 3.28) P = 0.255	174.25 vs. 168.45 5.80 (-1.91, 13.51) P = 0.134
<i>Current-threshold relation</i>		
Resting I/V-slope	0.593 vs. 0.559 6.1% (0.6%, 11.8%) P = 0.030	0.570 vs. 0.559 1.9% (-3.4%, 7.5%) P = 0.471
Minimum I/V-slope	0.258 vs. 0.238 8.5% (0.0%, 17.7%) P = 0.0498	0.232 vs. 0.238 -2.6% (-10.2%, 5.7%) P = 0.494
Hyperpolarizing I/V-slope	0.345 vs. 0.283 21.7% (3.5%, 43.0%) P = 0.019	0.309 vs. 0.283 9.2% (-7.0%, 28.3%) P = 0.271
<i>Recovery cycle</i>		
Refractoriness at 2 ms (%)	24.76 vs. 34.95 -10.192 (-17.160, -3.224) P = 0.006	34.00 vs. 34.95 -0.950 (-8.027, 6.127) P = 0.784
Superexcitability (%)	-27.78 vs. -26.41 -1.361 (-3.165, 0.442) P = 0.134	-25.52 vs. -26.41 0.889 (-0.914, 2.693) P = 0.322

Table 3 Continued on next page

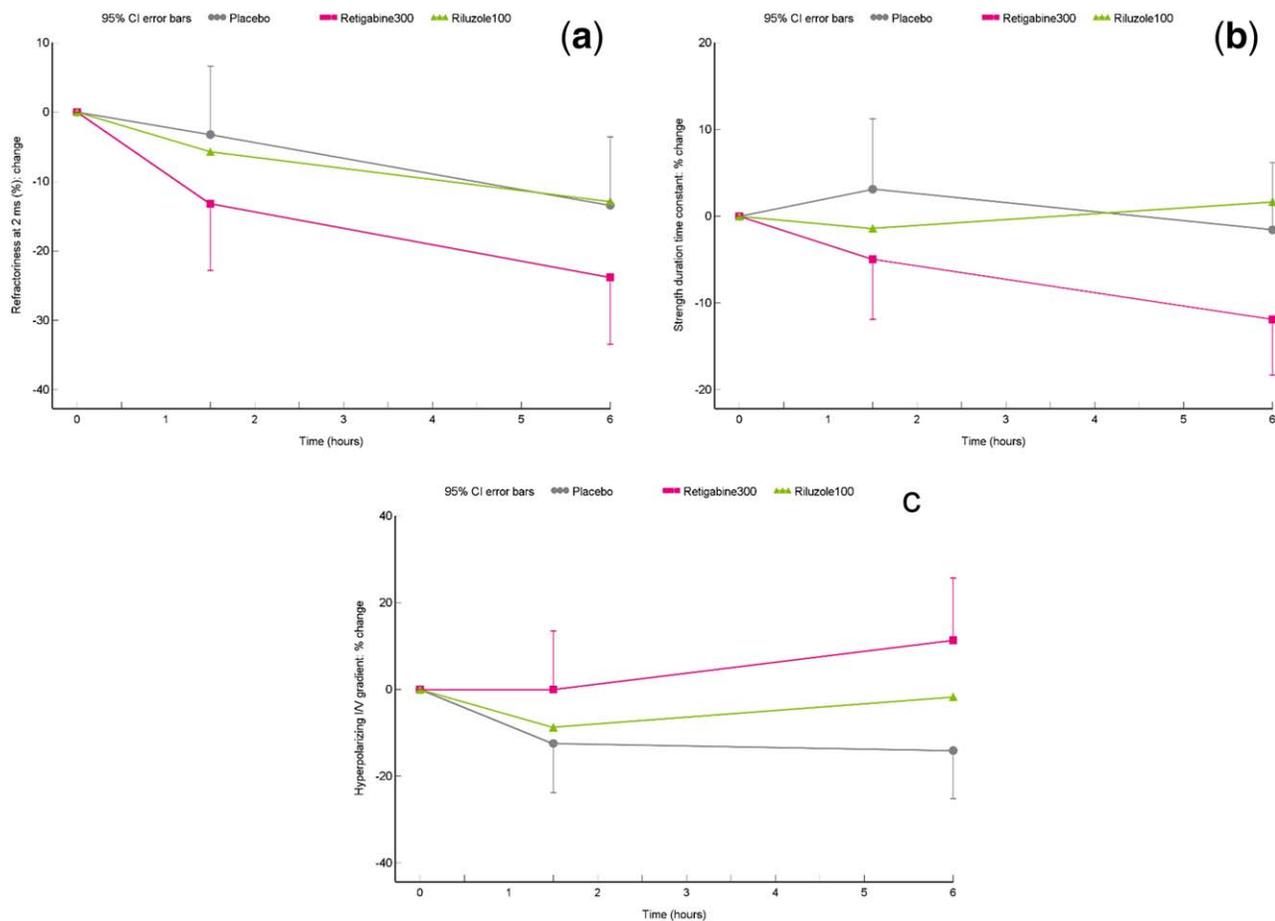
**Table 3 Continued**

Parameter	Contrast retigabine vs. placebo	Contrast riluzole vs. placebo
Subexcitability (%)	10.78 vs. 11.97 -1.196 (-2.715, 0.323) P = 0.117	12.03 vs. 11.97 0.059 (-1.472, 1.591) P = 0.937
Refractory period (ms)	2.41 vs. 2.58 -0.17 (-0.27, -0.06) P = 0.003	2.57 vs. 2.58 -0.01 (-0.12, 0.09) P = 0.788

Estimated mean of both postdose timepoints for each treatment vs. placebo. Treatment effects depicted as the estimated mean difference with placebo (95% CI of the difference).

In our study, a single dose of retigabine resulted in significant changes in various axonal excitability parameters. Strength duration properties showed a decrease in SDTC and increase in rheobase. Both can be explained by either hyperpolarization of resting membrane potential (since hyperpolarization decreases nodal persistent sodium current),<sup>24,25</sup> or a decrease in nodal electrical capacitance. As it is unlikely that retigabine induces histological changes resulting in decreased nodal capacitance, hyperpolarization of resting membrane potential is the most likely mechanism. Membrane hyperpolarization also results in increased threshold current needed to evoke a target CMAP of 50% of its maximum amplitude, similar to the findings in

previous studies.<sup>26</sup> In the current–threshold relationship, a steeper hyperpolarizing I/V slope was found, producing a larger inward rectification, which is also expected to be enhanced when the axonal membrane potential becomes more hyperpolarized. Furthermore, in the present study retigabine reduced refractoriness at 2 ms and the refractory period, which is consistent with a decrease in sodium-channel inactivation due to membrane hyperpolarization.<sup>27,28</sup> Shortening of the refractory period may also be due to early repolarization of the action potential. Early repolarization by retigabine has been observed previously as a result of membrane hyperpolarization induced by a hyperpolarizing shift of the voltage dependence of slow potassium channels.<sup>29</sup> This effect is most



**Figure 4** Change from baseline plot of the treatment effect on (a) refractoriness at 2 ms, (b) strength duration time constant, and (c) hyperpolarizing I/V slope. CI, confidence interval; Retigabine300, retigabine 300 mg treatment; Riluzole100, riluzole 100 mg treatment.

likely due to the greater number of potassium channels open at hyperpolarized membrane potentials. As riluzole has been previously shown to reduce refractoriness at 2 ms,<sup>13</sup> this observed effect of retigabine might be beneficial in ALS.

In the recovery cycle, retigabine did not induce significant changes in superexcitability and late subexcitability; this could be due to a plasma concentration being too low to induce detectable effects, as shown in a study with flupirtine.<sup>30</sup> In our study, we determined subexcitability after only a single supramaximal preconditioning pulse, where previous studies applied multiple preconditioning supramaximal pulses,<sup>26,31</sup> known to enhance late afterhyperpolarization, increasing subexcitability<sup>32</sup> and the sensitivity to detect treatment-induced changes.

Retigabine did not normalize any of the parameters that were found to be significantly different from healthy controls in our study. It did, however, change SDTC in the direction of normalization, a variable that has previously been shown to be abnormally increased in patients with ALS.<sup>2,4-6</sup> There was no significant correlation between retigabine concentration and effects on excitability variables (not shown); however, the appropriate approach would be to develop a population pharmacokinetic-pharmacodynamic model, which will be explored in the future. In any case, the chosen study design and statistical analysis ensure that the observed effects are induced by retigabine and cannot, for example, be explained by disease progression.

#### Effects of riluzole on peripheral nerve excitability

In our study, a single dose of riluzole had no significant effects on excitability variables. It is possible that the period of riluzole administration in our study was too short, since riluzole administration of, on average, 7 weeks in patients with ALS decreased refractoriness at 2 ms and superexcitability.<sup>13</sup> Nevertheless, we expected an effect on excitability variables after a single dose of riluzole, as it inhibits persistent sodium currents and shifts the voltage dependence of sodium-channel inactivation in a negative direction.<sup>33</sup> Another possible explanation of the lack of effect in our study may be related to the low riluzole concentrations that were still present during the baseline measurement, despite the preceding washout of standard riluzole treatment. This seems, however, unlikely because concentrations were ~5 times below the IC<sub>50</sub> for voltage-gated sodium channels and because no effect of riluzole on excitability variables was found in the riluzole-naïve patient of our study. Also, riluzole might have been less effective due to persistent sodium channels remaining in an open state.<sup>13</sup> Finally, there are indications that riluzole might lose its efficacy in later stages of the disease,<sup>34,35</sup> and our patients had been diagnosed and treated with riluzole, on average, more than a year prior to starting the study. A hypothesis for this loss of efficacy is that upregulation of efflux transporters in disease-affected regions,<sup>36</sup> such as P-glycoprotein (PGP) and breast cancer resistance protein (BCRP), would lead to very low concentrations at the target site.

#### Relation with disease progression

A relation between SDTC and ALSFRS-R<sup>8</sup> decline and SDTC and survival<sup>7</sup> has been reported, but in the current study no

correlation was observed between any of the excitability parameters and disease progression as measured by a change in ALSFRS-R. This could be due to the shorter follow-up (3 months compared to 6 months) and/or lower number of subjects in our study (18 vs. 60). When the follow-up was extended in our study in an *ad-hoc* analysis to, on average, 14.5 months (range 5.8–20.2 months) after baseline ALSFRS-R, again no significant correlation was found between excitability parameters and change in ALSFRS-R normalized for interval (time between baseline and second questionnaire) (data not shown).

#### Limitations

The most important limitation of our study was a lack of complete washout of standard riluzole treatment as described in the previous sections. As all but one participant were men, one should be cautious in extrapolating the results to female patients. Four (22%) of the enrolled patients had a familial history of ALS and therefore probably a hereditary form in which the pathophysiological mechanism might differ from that in patients with sporadic ALS.

#### Clinical implications

This study shows that a single dose of retigabine has a greater effect on peripheral nerve excitability than a single dose of riluzole, the current registered treatment for ALS. Previous studies showed that a prolonged SDTC is related to more rapid disease progression and shorter survival.<sup>7,8</sup> Although SDTC in our ALS patients was not statistically different from that in healthy controls, retigabine induced shortening of this variable. Long-term retigabine administration may, therefore, reverse the increased persistent sodium current underlying SDTC prolongation, which was suggested to induce hyperexcitability and motor neuron death.<sup>8,12</sup>

If, in the future, peripheral nerve excitability proves to be predictive of clinical outcome, it might be a very useful, noninvasive biomarker to test for potential treatments for ALS, and measure treatment efficacy on a much shorter basis compared to the sensitivity of the ALSFRS-R or survival measures.

## METHODS

### Subjects

Eighteen patients with ALS, aged between 18 and 80 years, were enrolled in the study, which was performed at the University Medical Center Utrecht in Utrecht, The Netherlands in collaboration with the Centre for Human Drug Research, Leiden, The Netherlands. Patients were recruited via the department's patient database, through advertisements, and the newsletters of the "Vereniging Spierziekten Nederland" (VSN), the organization for patients with neuromuscular disorders in the Netherlands. All patients gave written informed consent prior to any study-related activity, after which a screening visit evaluated eligibility. Main inclusion criteria were: mastery of the Dutch language, diagnosis of definite, probable, or probable laboratory-supported ALS according to the revised El Escorial criteria of 1998.<sup>37</sup> Fasciculations in the lower arm to be used for excitability measurements observed by the treating neurologist, CMAP of the abductor pollicis brevis muscle in the arm with fasciculations exceeding 1 mV, as well as no history of diabetes, neuropathy, or neuromuscular disorders other than ALS, carpal tunnel syndrome, trauma to the upper extremities, or other orthopedic conditions that might affect the electrophysiological measurements, and no medication that might affect electrophysiological

measurements, other than that used in the study. During the trial, the inclusion criterion for fasciculations was modified to include subjects with fasciculations anywhere in the arm, not only in the lower arm, and an exclusion for history of alcohol or drug dependence was removed. These changes were made in order to facilitate patient recruitment and were not considered to impact the study validity.

### Experimental design

We performed a randomized, double-blind, three-way crossover, placebo-controlled study of the test–retest reliability of peripheral motor nerve excitability and the effects of oral retigabine and riluzole on these measurements in patients with ALS. Visits were scheduled a week apart in order to allow a sufficiently long washout of riluzole and retigabine. Eligible subjects arrived at the research unit on the morning of a treatment visit, and after passing a brief re-eligibility and health check, they underwent baseline excitability testing. Capsules with medication were then swallowed with water on an empty stomach, after which regular blood samples were collected for pharmacokinetic analysis. Subjects were required to remain fasting until 2.5 hours after dosing, although water was allowed. At 1.5 and 6 hours after dosing, excitability testing was repeated. Blood samples for pharmacokinetic analysis were taken predose and 0.5, 1, 1.5, 2, 2.5, 3, 4, 6, and 7 hours after dosing. After the last blood sample the subject was discharged.

Since riluzole is the standard treatment for ALS patients (50 mg b.i.d.), we instituted a 1-day washout of riluzole before each dosing occasion to minimize the effect of the drug on excitability variables. Although the half-life of riluzole is ~9–15 hours<sup>38</sup> and not all of the drug was expected to have washed out of the system after 24 hours, it was deemed unethical to have a longer washout period, as this might have impacted the efficacy of the treatment, and the willingness of the patients to participate in the study.

The study was approved by the Independent Ethics Committee of the Foundation “Evaluation of Ethics in Biomedical Research” (Stichting Beoordeling Ethiek Biomedisch Onderzoek), Assen, The Netherlands. The study was registered in the Dutch Trial Registry (Nederlands Trial Register, NTR) under study number NTR6278 and was carried out between November 2015 and April 2017.

### Treatment, randomization, and masking

Subjects received a single dose of 100 mg ( $2 \times 50$  mg) riluzole (Rilutek, Sanofi, Paris, France), 300 mg (100 and 200 mg) retigabine (Trobalt, Glaxo Smith Kline, Brentford, UK) as encapsulated tablets or matching placebo capsules. Subjects were randomly assigned to a treatment order, with a balanced design using a code generated by an unblinded statistician who was otherwise not involved in the execution of the study. A masked physician enrolled patients into the study. Until study closure the treatment codes were only available to this statistician and the Leiden University Medical Center (LUMC) pharmacy, which distributed the study agents.

### Motor nerve excitability testing

Motor excitability was measured in the median nerve at the wrist. The setup consisted of Viking IV EMG apparatus (Nicolet Biomedical, Madison, WI), coupled to a computer (PCI-6221, National Instruments, Baltimore, MD) running QTRAC-S software (TRONDNF, v. 19-06-2015, Institute of Neurology, Queen Square, London, UK) and an isolated bipolar constant current stimulator (DS5, Digitimer, UK model D185-HB4). The median nerve was stimulated at the wrist via nonpolarizable surface electrodes (cathode at the wrist; anode 10 cm proximal over the radial side of the forearm). The thenar CMAP was recorded by surface electrodes in a belly-tendon montage. The distance between active recording electrode and stimulating cathode was 7 cm. The median nerve was warmed to 37°C by wrapping the forearm and hand for 30 minutes in a warm water blanket through which water at 37°C flowed constantly (Cincinnati Sub-zero Norm-O-Temp with Cincinnati Sub-zero maxi-therm lite infant hyper-hypothermia blanket for single patient use).<sup>39</sup>

During excitability testing the forearm and hand were kept in the blanket with flowing water at 37°C in order to maintain a constant nerve temperature. Skin temperature was continuously monitored by means of a sensor near the stimulating cathode. Distal motor latency (DML) was measured every 1.6 seconds during each excitability testing in order to check if changes in nerve temperature resulted in conduction changes. DML was defined as the point where the CMAP deviated by 10% of its amplitude from baseline to peak. These procedures were based on previous studies.<sup>39–41</sup>

To examine axonal excitability parameters, a specific sequence of conditioning and test stimuli were applied to the nerve. Conditioning stimuli were constant currents that either slightly depolarized resting membrane potential, slightly hyperpolarized resting membrane potential, or induced nerve action potentials. Threshold was defined as the test-stimulus current needed for a target CMAP of 40% of its maximum amplitude.

Each excitability test consisted of: stimulus response (SR) curve (relation between stimulus current and response amplitude), charge-duration ( $Q_t$ ) relation (relation between stimulus charge and stimulus duration), threshold electrotonus (time course of threshold changes during a depolarizing or hyperpolarizing conditioning current of 100 ms of 20% or 40% of the current for an unconditioned target response), I/V relation (relation between the magnitude of a 200 ms duration conditioning current, varying from 50% depolarizing to 100% hyperpolarizing, and the threshold at its end), and recovery cycle (time-course of the threshold changes after a supramaximal conditioning stimulus eliciting action potentials).

The following parameters were determined: threshold for an unconditioned target response of 50% (stimulus current required to evoke a CMAP of 50% of maximal), rheobase (slope of the  $Q_t$  relation), strength-duration time constant (SDTC; absolute value of the x-intercept of the  $Q_t$  relation), TE<sub>d</sub>90-100 (threshold decrease at the end of the 40% depolarizing conditioning stimulus), TE<sub>d</sub>40-60 (threshold decrease at 40–60 ms of the 40% depolarizing conditioning stimulus), TE<sub>d</sub>peak (maximal threshold decrease during 40% depolarizing conditioning stimulus), S<sub>2</sub>-accommodation (difference between TE<sub>d</sub>peak and TE<sub>d</sub>90-100), accommodation half-time (time between the onset of the conditioning stimulus and the timepoint where threshold decrease is halfway between TE<sub>d</sub>peak and TE<sub>d</sub>90-100), TE<sub>h</sub>90-100 (threshold increase at the end of the 40% hyperpolarizing conditioning stimulus), fanning (sum of the absolute values of TE<sub>d</sub>90-100 and TE<sub>h</sub>90-100), resting I/V slope (slope between –10% and +10% conditioning stimuli), minimal I/V slope (smallest slope in the hyperpolarizing part of the I/V curve), hyperpolarizing I/V-slope (slope between 100% and 80% hyperpolarizing conditioning stimuli), refractoriness at 2 ms (threshold change at the conditioning-test interval of 2 ms), refractory period (time between conditioning stimulus and return of threshold to baseline), superexcitability (lowest threshold after refractory period), and subexcitability (highest threshold after superexcitability).

Primary endpoints were repeatability of these variables as assessed by Cronbach’s alpha, and effects of riluzole and retigabine on the variables compared to placebo.

### Controls

In a separate study, we also investigated excitability by the same methods in 18 age-matched healthy controls (nine men, median age 53 years, range 35–71) who had no neurological symptoms and did not use medication.

### ALSFRS-R

The ALSFRS-R<sup>42</sup> evaluating disability in patients with ALS was performed at baseline and ~3 months after the first dose. This revised version of the ALSFRS, which incorporates additional assessments of dyspnea, orthopnea, and the need for ventilatory support, retains the properties of the original scale and shows strong internal consistency and construct validity. The rating scale is a validated, reliable, rating

instrument for monitoring the progression of disability in patients with ALS.<sup>43,44</sup>

### Data management

All data were stored in a clinical trial database (Promasys, Omnicomm, Fort Lauderdale, FL) and checked for accuracy and completeness. We performed a blinded data review before code-breaking and analysis according to a standard procedure at CHDR.

### Statistical analysis

Pharmacokinetic parameters, including  $C_{max}$  and  $T_{max}$ , were determined by standard noncompartmental methods using R software v. 3.4.0. Test-retest reliability of the primary endpoints was assessed by Cronbach's alpha, where a sample of 18 subjects was considered to be sufficient. An interim analysis evaluating between-day repeatability by Cronbach's alpha was performed after 12 patients had completed the study. A value of at least 0.7 (acceptable repeatability) was needed for at least one of the variables that were deemed most relevant—refractoriness at 2 ms, SDTC, superexcitability, TED40-60 or TED90-100—to proceed with the final six patients. To establish whether significant treatment effects could be detected on the excitability parameters, a mixed model analysis of covariance was used with treatment, time, and treatment by time as fixed factors and subject, subject by treatment, and subject by time as random factors, and the average baseline measurement as covariate. The Kenward-Rogers approximation was used to estimate denominator degrees of freedom and model parameters were estimated using the restricted maximum likelihood method. To determine treatment effects on excitability parameters the estimated averages of both postdose time-points for each treatment were compared with placebo. Missing data were not imputed, but were estimated within the statistical model. Residual Q-Q plots were used to check the assumption of normality of the error term in the mixed effects model together with the Shapiro-Wilk test for normality. Parameters violating the assumption of normality were log-transformed and after the analysis were backtransformed so that the results can be interpreted as a percentage change.

The excitability variables of ALS patients at the first, predose, visit were compared with those obtained in the healthy controls by unpaired Student's *t*-test.

The correlation between the difference in ALSFRS-R scores (at baseline and at 3 months) and the first visit predose excitability parameters was calculated using Spearman correlation. The significance level was set at  $P < 0.05$  and 95% confidence intervals of the estimated difference between the treatment and placebo groups are presented. All calculations were performed using SAS for windows v. 9.4 (SAS Institute, Cary, NC).

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The first two authors contributed equally to this work. The last two authors contributed equally to this work.

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### AUTHOR CONTRIBUTIONS

J.A.A.C.H. and M.K. wrote the article; J.A.A.C.H., M.K., B.T.H.M.S., L.vdB., T.A.F., H.F., and G.J.G. designed the research; J.A.A.C.H., M.K., and B.T.H.M.S. performed the research; J.A.A.C.H., M.K., B.T.H.M.S., and D.Z. analyzed the data.

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1. Bae, J.S., Simon, N.G., Menon, P., Vucic, S. & Kiernan, M.C. The puzzling case of hyperexcitability in amyotrophic lateral sclerosis. *J. Clin. Neurol.* **9**, 65–74 (2013).
2. Kanai, K. et al. Altered axonal excitability properties in amyotrophic lateral sclerosis: impaired potassium channel function related to disease stage. *Brain* **129**, 953–962 (2006).
3. Park, S.B., Kiernan, M.C. & Vucic, S. Axonal excitability in amyotrophic lateral sclerosis. *Neurotherapeutics* **14**, 78–90 (2017).
4. Vucic, S. & Kiernan, M.C. Axonal excitability properties in amyotrophic lateral sclerosis. *Clin. Neurophysiol.* **117**, 1458–1466 (2006).
5. Geevasinga, N., Menon, P., Howells, J., Nicholson, G.A., Kiernan, M.C. & Vucic, S. Axonal ion channel dysfunction in c9orf72 familial amyotrophic lateral sclerosis. *JAMA Neurol.* **72**, 49–57 (2015).
6. Menon, P., Kiernan, M.C. & Vucic, S. ALS pathophysiology: insights from the split-hand phenomenon. *Clin. Neurophysiol.* **125**, 186–193 (2014).
7. Kanai, K. et al. Motor axonal excitability properties are strong predictors for survival in amyotrophic lateral sclerosis. *J. Neurol. Neurosurg. Psychiatry* **83**, 734–738 (2012).
8. Shibuya, K. et al. Increased motor axonal persistent sodium currents predict rapid functional declines in amyotrophic lateral sclerosis. *Neural. Clin. Neurosci.* **4**, 108–111 (2016).
9. Krarup, C. Lower motor neuron involvement examined by quantitative electromyography in amyotrophic lateral sclerosis. *Clin. Neurophysiol.* **122**, 414–422 (2011).
10. Kuwabara, S. & Misawa, S. Axonal ionic pathophysiology in human peripheral neuropathy and motor neuron disease. *Curr. Neurovasc. Res.* **1**, 373–379 (2004).
11. de Carvalho, M. Why is ALS so excited? *Clin. Neurophysiol.* **122**, 1689–1690 (2011).
12. Wainger, B.J. et al. Intrinsic membrane hyperexcitability of amyotrophic lateral sclerosis patient-derived motor neurons. *Cell. Rep.* **7**, 1–11 (2014).
13. Vucic, S. et al. Riluzole exerts central and peripheral modulating effects in amyotrophic lateral sclerosis. *Brain* **136**, 1361–1370 (2013).
14. Tomlinson, S.E., Tan, S.V., Kullmann, D.M., Burke, D., Hanna, M.G. & Bostock, H. Axonal excitability changes in genetic neuronal ion channel disorders. *J. Peripher. Nerv. Syst.* **14**, 144–145 (2009).
15. Agency, E.M. SmPC Rilutek. <[http://www.ema.europa.eu/docs/en\\_GB/document\\_library/EPAR\\_-\\_Product\\_Information/human/000109/WC500056586.pdf](http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Product_Information/human/000109/WC500056586.pdf)> (2018).
16. Gunthorpe, M.J., Large, C.H. & Sankar, R. The mechanism of action of retigabine (ezogabine), a first-in-class K<sup>+</sup> channel opener for the treatment of epilepsy. *Epilepsia* **53**, 412–424 (2012).
17. Huang, C.J. et al. Characterization of voltage-gated sodium-channel blockers by electrical stimulation and fluorescence detection of membrane potential. *Nat. Biotechnol.* **24**, 439–446 (2006).
18. Bostock, H. & Baker, M. Evidence for two types of potassium channel in human motor axons in vivo. *Brain Res.* **462**, 354–358 (1988).
19. Reid, G., Scholz, A., Bostock, H. & Vogel, W. Human axons contain at least five types of voltage-dependent potassium channel. *J. Physiol.* **518** (Pt 3), 681–696 (1999).
20. Schwarz, J.R. et al. KCNQ channels mediate IKs, a slow K<sup>+</sup> current regulating excitability in the rat node of Ranvier. *J. Physiol.* **573**, 17–34 (2006).
21. Devaux, J.J., Kleopa, K.A., Cooper, E.C. & Scherer, S.S. KCNQ2 is a nodal K<sup>+</sup> channel. *J. Neurosci.* **24**, 1236–1244 (2004).
22. Tatulian, L., Delmas, P., Abogadie, F.C. & Brown, D.A. Activation of expressed KCNQ potassium currents and native neuronal M-type

- potassium currents by the anti-convulsant drug retigabine. *J. Neurosci.* **21**, 5535–5545 (2001).
23. Wua, Y.J. & Dworetzky, S.I. Recent developments on KCNQ potassium channel openers. *Curr. Med. Chem.* **12**, 453–460 (2005).
  24. Kiernan, M.C. & Bostock, H. Effects of membrane polarization and ischaemia on the excitability properties of human motor axons. *Brain* **123** (Pt 12), 2542–2551 (2000).
  25. Bostock, H. & Rothwell, J.C. Latent addition in motor and sensory fibres of human peripheral nerve. *J. Physiol.* **498** (Pt 1), 277–294 (1997).
  26. Lang, P.M., Fleckenstein, J., Passmore, G.M., Brown, D.A. & Grafe, P. Retigabine reduces the excitability of unmyelinated peripheral human axons. *Neuropharmacology* **54**, 1271–1278 (2008).
  27. Kiernan, M.C., Guglielmi, J.M., Kaji, R., Murray, N.M. & Bostock, H. Evidence for axonal membrane hyperpolarization in multifocal motor neuropathy with conduction block. *Brain* **125**, 664–675 (2002).
  28. Kiernan, M.C., Mogyoros, I. & Burke, D. Differences in the recovery of excitability in sensory and motor axons of human median nerve. *Brain* **119** (Pt 4), 1099–1105 (1996).
  29. Corbin-Leftwich, A., Mossadeq, S.M., Ha, J., Ruchala, I., Le, A.H. & Villalba-Galea, C.A. Retigabine holds KV7 channels open and stabilizes the resting potential. *J. Gen. Physiol.* **147**, 229–241 (2016).
  30. Fleckenstein, J., Sittl, R., Averbek, B., Lang, P.M., Imich, D. & Carr, R.W. Activation of axonal Kv7 channels in human peripheral nerve by flupirtine but not placebo — therapeutic potential for peripheral neuropathies: results of a randomised controlled trial. *J. Transl. Med.* **11**, 34 (2013).
  31. Sittl, R., Carr, R.W., Schwarz, J.R. & Grafe, P. The Kv7 potassium channel activator flupirtine affects clinical excitability parameters of myelinated axons in isolated rat sural nerve. *J. Peripher. Nerv. Syst.* **15**, 63–72 (2010).
  32. Baker, M., Bostock, H., Grafe, P. & Martius, P. Function and distribution of three types of rectifying channel in rat spinal root myelinated axons. *J. Physiol.* **383**, 45–67 (1987).
  33. Bellingham, M.C. A review of the neural mechanisms of action and clinical efficiency of riluzole in treating amyotrophic lateral sclerosis: what have we learned in the last decade? *CNS Neurosci. Ther.* **17**, 4–31 (2011).
  34. Zoccolella, S. *et al.* Riluzole and amyotrophic lateral sclerosis survival: a population-based study in southern Italy. *Eur. J. Neurol.* **14**, 262–268 (2007).
  35. Schuster, J.E., Fu, R., Siddique, T. & Heckman, C.J. Effect of prolonged riluzole exposure on cultured motoneurons in a mouse model of ALS. *J. Neurophysiol.* **107**, 484–492 (2012).
  36. Jablonski, M.R. *et al.* Selective increase of two ABC drug efflux transporters at the blood-spinal cord barrier suggests induced pharmacoresistance in ALS. *Neurobiol. Dis.* **47**, 194–200 (2012).
  37. Brooks, B.R., Miller, R.G., Swash, M., Munsat, T.L. & World Federation of Neurology Research Group on Motor Neuron, D. El Escorial revisited: revised criteria for the diagnosis of amyotrophic lateral sclerosis. *Amyotroph. Lateral Scler. Other Motor Neuron Disord.* **1**, 293–299 (2000).
  38. Le, L.A. *et al.* Single- and multiple-dose pharmacokinetics of riluzole in white subjects. *J. Clin. Pharmacol.* **37**, 820–827 (1997).
  39. Drenthen, J., Blok, J.H., van Heel, E.B. & Visser, G.H. Limb temperature and nerve conduction velocity during warming with hot water blankets. *J. Clin. Neurophysiol.* **25**, 104–110 (2008).
  40. Franssen, H. & Wieneke, G.H. Nerve conduction and temperature: necessary warming time. *Muscle Nerve* **17**, 336–344 (1994).
  41. Geerlings, A.H. & Mechelse, K. Temperature and nerve conduction velocity, some practical problems. *Electromyogr. Clin. Neurophysiol.* **25**, 253–259 (1985).
  42. Cedarbaum, J.M. *et al.* The ALSFRS-R: a revised ALS functional rating scale that incorporates assessments of respiratory function. BDNF ALS Study Group (Phase III). *J. Neurol. Sci.* **169**, 13–21 (1999).
  43. Kaufmann, P. *et al.* Excellent inter-rater, intra-rater, and telephone-administered reliability of the ALSFRS-R in a multicenter clinical trial. *Amyotroph. Lateral Scler.* **8**, 42–46 (2007).
  44. Kollwe, K., Mauss, U., Krampfl, K., Petri, S., Dengler, R. & Mohammadi, B. ALSFRS-R score and its ratio: a useful predictor for ALS-progression. *J. Neurol. Sci.* **275**, 69–73 (2008).